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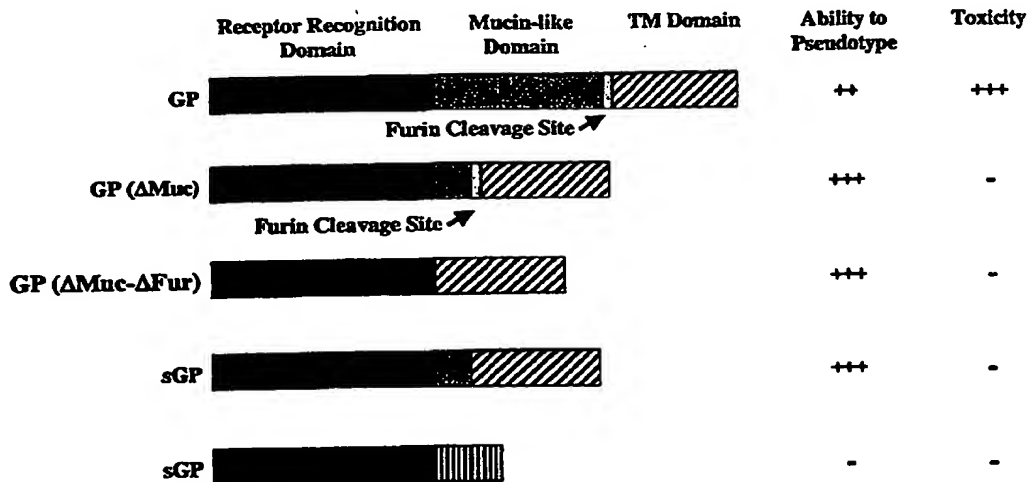
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<table style="width: 100%;"> <tr> <td style="width: 50%; vertical-align: top;"> <p>(21) International Application Number: PCT/US99/01382</p> <p>(22) International Filing Date: 21 January 1999 (21.01.99)</p> <p>(30) Priority Data: 60/072,033 21 January 1998 (21.01.98) US</p> <p>(63) Related by Continuation (CON) or Continuation-in-Part (CIP) to Earlier Application US 60/072,033 (CON) Filed on 21 January 1998 (21.01.98)</p> <p>(71) Applicant (for all designated States except US): THE REGENTS OF THE UNIVERSITY OF MICHIGAN [US/US]; Technology Management Office, Wolverine Tower, Room 2071, 3003 South State Street, Ann Arbor, MI 48109-1280 (US).</p> <p>(72) Inventors; and (75) Inventors/Applicants (for US only): NABEL, Gary, J. [US/US]; 385 Meadow Creek Drive, Ann Arbor, MI 48105 (US). DELGADO, Rafael [ES/US]; 1265 Crescent, Ann Arbor, MI 48103 (US). YANG, Zhi-yong [CN/US]; 2863 Leslie Park Circle, Ann Arbor, MI 48105 (US).</p> </td> <td style="width: 50%; vertical-align: top;"> <p>(74) Agents: SMITH, DeAnn, F. et al.; Harness, Dickey & Pierce, P.L.C., P.O.Box 828, Bloomfield Hills, MI 48303 (US).</p> <p>(81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, GH, GM, HR, HU, ID, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).</p> <p>Published With international search report.</p> </td> </tr> </table>			<p>(21) International Application Number: PCT/US99/01382</p> <p>(22) International Filing Date: 21 January 1999 (21.01.99)</p> <p>(30) Priority Data: 60/072,033 21 January 1998 (21.01.98) US</p> <p>(63) Related by Continuation (CON) or Continuation-in-Part (CIP) to Earlier Application US 60/072,033 (CON) Filed on 21 January 1998 (21.01.98)</p> <p>(71) Applicant (for all designated States except US): THE REGENTS OF THE UNIVERSITY OF MICHIGAN [US/US]; Technology Management Office, Wolverine Tower, Room 2071, 3003 South State Street, Ann Arbor, MI 48109-1280 (US).</p> <p>(72) Inventors; and (75) Inventors/Applicants (for US only): NABEL, Gary, J. [US/US]; 385 Meadow Creek Drive, Ann Arbor, MI 48105 (US). DELGADO, Rafael [ES/US]; 1265 Crescent, Ann Arbor, MI 48103 (US). YANG, Zhi-yong [CN/US]; 2863 Leslie Park Circle, Ann Arbor, MI 48105 (US).</p>	<p>(74) Agents: SMITH, DeAnn, F. et al.; Harness, Dickey & Pierce, P.L.C., P.O.Box 828, Bloomfield Hills, MI 48303 (US).</p> <p>(81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, GH, GM, HR, HU, ID, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).</p> <p>Published With international search report.</p>
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(54) Title: TARGETING GENE TRANSFER VECTORS TO CERTAIN CELL TYPES BY PSEUDOTYPING WITH VIRAL GLYCOPROTEIN



(57) Abstract

The present invention provides compositions and methods for targeting gene transfer vectors to certain cell types by pseudotyping with a transmembrane form of viral glycoprotein, such as that from Ebola virus. The methods comprise the step of administering to a cell population a gene to be transferred operatively linked to an appropriate transfer vehicle, wherein the transfer vehicle is associated with a transmembrane form of viral glycoprotein.

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TARGETING GENE TRANSFER VECTORS TO CERTAIN CELL TYPES BY PSEUDOTYPING WITH VIRAL GLYCOPROTEIN

FIELD OF THE INVENTION

5 The present invention relates generally to compositions and methods for selective gene transfer, and in particular, to methods for targeting genes to certain cell types, comprising introducing to a cell population the gene to be transferred operatively-linked to an appropriate transfer vehicle, wherein the transfer vehicle is associated with a transmembrane form of viral glycoprotein.

BACKGROUND OF THE INVENTION

10 Ebola virus has been identified as the cause of several highly lethal outbreaks of hemorrhagic fever. Infection begins typically with flu-like symptoms which often progress rapidly to fatal complications of hemorrhage, fever, and hypotensive shock. Bowen, E.T.W. et al., *Lancet* 1:571 (1977); Centers for Disease Control, *M.M.W.R.* 15 44:381 (1995); Le Guenno, B. et al., *Lancet* 345:1271 (1995); Peters, C.J. et al., *Fields Virology*, B.N. Fields, D.M. Knipe and P.M. Howley, Eds. (Lippincott-Raven, Philadelphia) p. 1161 (1996). The negative-stranded genome of Ebola virus contains seven structural and regulatory proteins (Sanchez, A. et al., *Virus Res.* 29:215 (1993)), but despite its relative simplicity, the molecular basis for Ebola virus 20 pathogenicity is unknown. Among the viral gene products, the glycoprotein is found in two forms: a secreted form, 50-70 kD (Sanchez, A. et al., *PNAS (USA)* 93:3602 (1996)), synthesized at high levels early in the course of infection, and an alternative transmembrane form, which arises from RNA editing to encode a 120-150 kD glycoprotein that is incorporated into the virion. Sanchez, A. et al., *PNAS (USA)* 25 93:3602 (1996); Volchkov, V.E. et al., *Virology* 214:421 (1995). The first 295 amino acids (aa) of both proteins are identical in the Zaire strain, while sGP contains an additional 69 and GP another 381 COOH- terminal aa residues. Sanchez, A. et al., *PNAS (USA)* 93:3602 (1996). The specific cellular targets of these related gene products and their roles in the pathogenesis of Ebola infection have not been 30 characterized.

SUMMARY OF THE INVENTION

 The present invention provides compositions and methods for targeting gene transfer vectors to certain cell types by pseudotyping with a transmembrane form of viral glycoprotein. In one embodiment, the methods of the invention comprise the

- 2 -

step of administering to a cell population a gene to be transferred operatively-linked to an appropriate transfer vehicle, wherein the transfer vehicle is associated with a transmembrane form of Ebola glycoprotein. In this embodiment, the gene will be targeted to cell types naturally infected with Ebola such as endothelial cells, monocytes and hepatocytes.

Genetic constructs for selective gene transfer into certain cell types are also provided. The genetic constructs of the present invention comprise a gene to be transferred operatively-linked to an appropriate transfer vehicle or carrier, wherein the transfer vehicle or carrier is associated with a transmembrane form of viral glycoprotein. In one embodiment, the transmembrane form of Ebola glycoprotein is expressed on the surface of a virus-based gene-targeting vector, *e.g.*, lentiviral or retroviral vector. In another embodiment, an expressed or synthesized transmembrane glycoprotein is chemically derivatized to a non-biologic gene targeting vehicle.

Additional objects, advantages, and features of the present invention will become apparent from the following description and appended claims, taken in conjunction with the accompanying drawings.

BRIEF DESCRIPTION OF THE DRAWINGS

The various advantages of the present invention will become apparent to one skilled in the art by reading the following specification and subjoined claims and by referencing the following drawings.

Figures 1A-1C show the binding of sGP to neutrophils;

Figures 2A-2D show the infection of different cell types by a GP-pseudotyped vector of the present invention;

Figures 3A-3F show the dependence of sGP binding on CD16b and correlation of binding with neutrophil activation;

Figures 4A-4B show the effect of sGP on neutrophil function;

Figures 5A-5C show the infection rate of cells with a GP-pseudotyped retroviral vector of the present invention;

Figure 6 is a schematic of the plasmid pVR 1012-GP(IC) (Ivory Coast strain of GP, see SEQ ID NO: 1);

Figure 7 is a schematic of the plasmid pVR 1012-GP(S) (Sudan strain of GP, see SEQ ID NO: 2);

Figure 8 is a schematic of the plasmid pVR 1012-GP(Z) (Zaire strain of GP, see SEQ ID NO: 3);

Figure 9 is a schematic of the plasmid pVR 1012-sGP(Z) (Zaire strain of sGP, see SEQ ID NO: 4); and

Figure 10 is a summary of the characterization of GP and sGP derivatives for their ability to pseudotype to induce cytotoxicity in producer cells.

5 DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

The present invention provides genetic constructs and methods for targeting gene transfer vectors to certain cell types by pseudotyping with a transmembrane form of viral glycoprotein. The methods for selective gene transfer of the present invention comprise the step of administering to a cell population a genetic construct
10 of the present invention so that the gene is transferred and expressed in certain cell types present in the cell population. Administration to the cell population may be *ex vivo* or *in vivo*.

The genetic constructs of the present invention comprise a gene to be transferred operatively-linked to an appropriate transfer vehicle or carrier, wherein the
15 transfer vehicle or carrier is associated with a transmembrane form of viral glycoprotein. In one embodiment, the transmembrane form of Ebola glycoprotein is associated with the vehicle or carrier. The gene to be transferred will thus be targeted to cell types naturally infected with Ebola virus including endothelial cells, hepatocytes, monocytes and related cell types such as dendritic cells. The
20 transmembrane form of Ebola glycoprotein may be chosen from, without limitation, the Ivory Coast strain (SEQ ID NO: 1), Sudan strain (SEQ ID NO: 2), the Zaire strain (SEQ ID NO: 3) and/or the Reston strain. It will be appreciated that in other embodiments of the present invention, other hemorrhagic fever virus glycoproteins, in particular transmembrane glycoproteins, may be employed and will target those cell
25 types naturally infected by the virus. Examples of hemorrhagic viruses include dengue virus, Yellow Fever virus (*flaviviridae*); Lassa, Junin and Machupo (*arenaviridae*); Rift Valley, Congo-Crimean and Hantaan (*bunyaviridae*); and Marburg (*filoviridae*). It will also be appreciated that derivatives of the transmembrane glycoprotein which retain the capability of targeting specific cell types, may also be
30 employed, for example, the transmembrane glycoproteins may be mutated, e.g., toxic regions may be removed to improve producer cell viability (see Figure 10).

The transmembrane glycoprotein may be expressed on the surface of a virus-based gene-targeting vector, e.g., lentiviral, retroviral, replication-deficient retroviral, adenoviral or adeno-associated viral vector. The transmembrane glycoprotein may

- 4 -

also be expressed or synthesized and chemically derivatized to a non-biologic gene targeting vehicle, e.g., liposome or DNA-protein complex.

The term "operatively-linked" as used herein refers to functional linkage between a nucleic acid expression control sequence (such as a promoter) and a second nucleic acid sequence (*i.e.*, gene), wherein the expression control sequence directs transcription of the nucleic acid corresponding to the second sequence. Expression control sequences are known to those skilled in the art (see, e.g., Goeddel, *Gene Expression Technology: Methods in Enzymology* 185, Academic Press, San Diego, CA (1990)). "Associated with" as used herein refers to the transmembrane form of viral glycoprotein being in contact or linkage with the transfer vehicle or carrier in such a way as to direct the transfer vehicle or carrier to certain cell types. The terms "transfer vehicle" and "carrier" refer to any type of structure which is capable of delivering the gene of interest to a target cell.

Many transfer vehicles or carriers are known in the art. For example, various viruses that are capable of infecting cells can be recombinantly manipulated to carry the gene of interest without affecting their infectivity. As used herein, the terms "infect" and "infectivity" refer only to the ability of a virus to transfer genetic material to a target cell. Those terms do not mean that the virus is capable of replication in the target cell. In fact, it is preferable that such viruses are replication defective so that target cells do not suffer the effects of viral replication.

In one embodiment, the virus employed is a replication defective retroviruses. When these replication defective retroviruses are employed, their genomes can be packaged by a helper virus in accordance with well-known techniques. Suitable retroviruses include PLJ, pZip, pWe and pEM, each of which is well known in the art. Suitable helper viruses for packaging genomes include ψ Crip, ψ Cre, ψ 2, ψ Am and adeno-associated viruses.

In another embodiment, lentiviral vectors are employed. Surprisingly, the inventors of the present invention were successful in pseudotyping lentiviral vectors (HIV) with the transmembrane glycoprotein from Ebola. Feline immunodeficiency virus, bovine immunodeficiency virus, simian immunodeficiency virus and EAIIV, may also be employed as the carrier in the compositions and methods of the present invention.

Gene delivery systems other than viruses may also be employed. For example, the gene to be transferred may be packaged in a liposome which is chemically derivatized to the transmembrane glycoprotein. To form these liposomes,

- 5 -

one mixes the DNA of an expression vector which expresses the gene to be transferred with lipid, such as *N*-[1-(2,3,-dioleoyloxy)propyl]-*N,N,N*-trimethylammonium chloride (DOTMA) in a suitable buffer, such as Hepes buffered saline. This causes the spontaneous formation of lipid-DNA complexes (liposomes). Felgner, P.L. et al.,
5 *PNAS (USA)* 84:7413-7417 (1987).

Another gene delivery system that may be utilized in this invention is DNA-protein complexes. The formation of DNA-protein complexes is described in United States Patent No. 5,166,320, the disclosure of which is herein incorporated by reference.

10 It will be appreciated that any gene may be employed in the compositions and methods of the present invention. For example, and without limitation, in the treatment of cancer, death inducing genes, including genes coding for cytostatic or cytotoxic proteins, e.g., HSV tk, and genes encoding cyclin dependent kinase inhibitors, p21, p27, cytosine deaminase, and fas ligand, may all be employed. In
15 another example, for the treatment of cardiovascular or ischemic vascular disease, genes encoding angiogenic factors such as VEGF basic or acidic FGF's (FGF 1-5) may be employed. In yet another example, in the treatment of vasospasm, the gene encoding NO synthase or heme oxygenase, may be employed. In a further example, monocytes and dendritic cells may be targeted with genes encoding immunogens for
20 cell-targeted immunization.

In one embodiment, the methods of targeting gene transfer vectors to certain cell types involve administering to a cell population *ex vivo*, a construct of the present invention and introducing the transfected cells into a subject. In an alternative
25 embodiment, the methods of the present invention comprise administering to an *in vivo* cell population a construct of the present invention. Administration can be by any of the routes normally used for *in vivo* gene therapy such as direct delivery to cells via a gene gun, and other known techniques. The constructs are thus administered in any suitable manner, preferably with pharmaceutically acceptable carriers. The constructs can be administered, for example, by intravenous infusion, orally, topically,
30 intraperitoneally, intravesically or intrathecally. The preferred method of administration will often be intravenous.

To practice an *ex vivo* method of the present invention, a source of cells is obtained. The cells are optionally selected from *in vitro* cells, such as those derived from cell culture and *ex vivo* cells, such as those derived from a subject. The term
35 "subject" is intended to include living organisms, e.g., mammals. Examples of

- 6 -

subjects include humans, primates, dogs, cats, mice, rats, and transgenic species thereof. It will be appreciated that specific cell populations may be obtained by isolation from certain tissues by methods known to those skilled in the art. The cells are maintained under conditions necessary to support growth, for example an appropriate temperature (e.g., 37°C) and atmosphere (e.g., air plus 5% CO₂).

The cells are then transfected with the constructs of the present invention by introducing the constructs to the cell population, under conditions favorable for transfection. According to one embodiment of the present invention, cells are treated with compounds that facilitate uptake of the constructs by the cells. According to another embodiment of the present invention, cells are treated with compounds that stimulate cell division and facilitate uptake of the constructs. It will be appreciated that compounds that facilitate uptake of constructs by cells and compounds that stimulate cell division are known to those skilled in the art.

The constructs of the present invention express the transferred gene in a dose dependent manner. The specific dose to be administered to a patient will be determined by the efficacy of the particular construct and/or delivery system employed, the gene transferred, and the condition of the patient, as well as the body weight or surface area of the patient to be treated. The size of the dose also will be determined by the existence, nature, and extent of any adverse side-effects that accompany the administration of a particular construct or effect a particular patient. In determining the effective amount of the construct or transfected cell to be administered, the physician needs to evaluate circulating plasma levels, toxicities, and progression of disease. It will be appreciated that administration can be accomplished via single or divided doses.

There is a wide variety of suitable formulations for pharmaceutical compositions containing the constructs of the present invention. Formulations suitable for oral administration can consist of (a) liquid solutions, such as an effective amount of the construct dissolved in diluents, such as water, saline or PEG 400; (b) capsules, sachets or tablets, each containing a predetermined amount of the construct, as liquids, solids, granules or gelatin; (c) suspensions in an appropriate liquid; and (d) suitable emulsions. The construct, alone or in combination with other suitable components, may also be made into aerosol formulations to be administered via inhalation, e.g., to the bronchial passageways. Aerosol formulations can be placed into pressurized acceptable propellants, such as dichlorodifluoromethane, propane, nitrogen, and the like.

Suitable formulations for rectal administration include, for example, suppositories, which consist of the construct with a suppository base. Suitable suppository bases include natural or synthetic triglycerides or paraffin hydrocarbons. In addition, it is also possible to use gelatin rectal capsules which consist of a combination of the construct with a base, including, for example, liquid triglycerides, polyethylene glycols, and paraffin hydrocarbons.

Formulations suitable for parenteral administration, such as, for example, by intraarticular (in the joints), intravenous, intramuscular, intradermal, intraperitoneal, and subcutaneous routes, include aqueous and non-aqueous, isotonic sterile injection solutions, which contain antioxidants, buffers, bacteriostats, and solutes that render the formulation isotonic with the blood of the intended recipient, and aqueous and non-aqueous sterile suspensions that can include suspending agents, solubilizers, thickening agents, stabilizers, and preservatives. The formulations can be presented in unit-dose or multi-dose sealed containers, such as ampules or vials. Extemporaneous injection solutions and suspensions can be prepared from sterile powders, granules, and tablets of the kind previously described. Cells transfected by the constructs as described above in the context of *ex vivo* therapy can also be administered as described above.

This invention also provides compositions and kits comprising the constructs of the present invention. For example, the composition can comprise the constructs of the present invention in a pharmaceutically acceptable carrier as described above. Kits comprising such compositions and instructions for use are also within the scope of this invention.

In order to more fully demonstrate the advantages arising from the present invention, the following examples are set forth. It is to be understood that the following is by way of example only and is not intended as a limitation on the scope of the invention.

SPECIFIC EXAMPLE 1

I. Methods

Recombinant retroviruses were produced by transient transfection of 293T cells: 2×10^6 cells were plated 24 hours before transfection in 60 mm dishes. Transfection was performed by calcium-phosphate precipitation using 3 μ g of a retroviral vector (Kinsella, T.M. et al., *Hum. Gene Ther.* 7:1405 (1996)) encoding luciferase linked to an internal ribosome entry site and a green fluorescent protein derivative (GFP; pEGFP, Clontech), pLZR_s-Luc-Gfp, 5 μ g of an expression vector

- 8 -

encoding gag and pol, pNGVL-MLVgag-pol, and 1 μ g of the envelope encoding plasmid: pNGVL-4070A (ampho) env, pCMV-Eco env or p1012-Ebola GP, respectively. Supernatants corresponding to 24-48 hours post-transfection were harvested, cleared by low-speed centrifugation and either used immediately for infection or frozen at -80°C. Infections were performed in 6-well plates ($1.5-2.5 \times 10^5$ adherent cells) or 12-well plates (5×10^5 non-adherent) using different dilutions of the supernatants by incubating the cells overnight with 1 ml and 300 μ l, respectively of the diluted supernatants. Polybrene was used at a concentration of 5 μ g/ml for all the cell lines except for D17 in which the concentration was 100 μ g/ml. After overnight infection, fresh medium was added and the cells were incubated for an additional 24 hours. After infection, the cells were lysed in 25 mM Tris-phosphate pH 8, 2 mM DTT, 2 mM 1,2-diaminocyclohexene-N,N,N',N'-tetraacetic acid, 10% glycerol, 1% TritonX-100, and assayed for luciferase activity using Luciferase Assay Reagent (Promega, Madison, WI) in a 1251 BioOrbit Luminometer. The same number of cells (range $5-10 \times 10^4$) was analyzed for every specific cell line.

Binding of sGP to neutrophils and inverse correlation of binding with activation: Figures 1A-1A2. PBMC from normal volunteers were incubated with control or sGP supernatants derived from transfected 293 cells, and immunostaining was performed using a rabbit antibody to sGP as previously described. Sanchez, A. et al., *PNAS (USA)* 93:3602 (1996); Xu, L. et al., *Nat. Med.* (1997) in press. Secondary staining was performed with a fluorescein isothiocyanate (FITC)-conjugated goat anti-rabbit IgG antibody (Sigma, F9887). All incubations were performed at 4°C for 30 minutes with .4 μ g of the relevant antibodies per 10^6 cells in a 50 μ l volume.

Figures 1B-1B1. Double immunostaining with antibodies to sGP and the neutrophil-specific marker, CD15. Cells were incubated with a FITC conjugated mouse anti-human CD15 antibody (Caltag, cat# MHCD1501), followed by secondary staining with a PE-conjugated anti-rabbit IgG antibody (Sigma) to detect sGP binding. Cells were washed with PBS, fixed in 1% formaldehyde, and analyzed by FACS.

Figure 1C. Specific absorption of sGP by neutrophils. Control or sGP supernatants derived from relevant transfected 293 cells (Xu, L. et al., *Nat. Med.* (1997) in press) were incubated at 1:500 dilution with 10^6 mononuclear or granulocytic cells. Cells were removed and the resulting supernatants analyzed by an 8% SDS PAGE gel. Western blot analysis was performed as previously described (Xu, L. et al., *Nat. Med.* (1997) in press) using an anti-GP rabbit antisera and a secondary

- 9 -

antibody, horseradish peroxidase conjugated donkey anti-rabbit IgG at a dilution of 1:5,000 (Amersham, NA934). Primary antibody was incubated for 30 minutes at room temperature, as was the secondary antibody. The immunocomplexes were detected by chemiluminescence using Supersignal® chemiluminescent substrate reagents (Pierce) according to the manufacturer's instructions. Arrow indicates sGP reactive band.

Infection of different cell types by GP-pseudotyped retroviral vector and preferential binding to endothelial cells: Figure 2A.

Infection of different indicator cell lines with the Ebola-GP pseudotyped retrovirus expressing luciferase. Amphotropic and ecotropic retroviral vectors were used as controls. Viruses were diluted to different multiplicities of infection (MOI) to provide for equal luciferase activity on Hela cervical epithelial cells, permissive for amphotropic retrovirus, D17 dog osteosarcoma cells (Embretson, J.E. et al., *J. Virol.* 61:3454 (1987)), which are permissive for amphotropic, xenotropic, and ecotropic retroviruses, and BW5147 T leukemia cells permissive for amphotropic and ecotropic virus. In these groups, GP virus titer was $1-4 \times 10^5$ /ml and amphotropic virus was $\sim 2 \times 10^4$ /ml (MOI's ≈ 1.0 and 0.1 , respectively), and the ecotropic virus titer was $\sim 10^6$ /ml (MOI ≈ 10). Titers were determined by endpoint dilution of reporter activity of the amphotropic virus in D17 cells, and was normalized to reverse transcriptase activity for the GP virus.

Figure 2B. Analysis of different normal or transformed cell lines by infection with amphotropic or GP retroviral vectors at the same titer (10^4 /ml, MOI ≈ 0.2). Forty-eight hours after infection, an equivalent of 5×10^4 cells was assayed for luciferase activity after exposure to equal titers of viral stocks. Luminescence is expressed as the fold-increase over non-infected control cells.

Figures 2C-2C3. The binding of sGP (left) or GP-pseudotyped retrovirus (right) to neutrophils (upper panel) or microvascular endothelium (lower panel) was determined by FACS. sGP binding was performed as in Fig. 1A, and retrovirus incubation was performed at 37°C for 2 hours in the presence of polybrene ($8 \mu\text{g}/\text{ml}$).

Figure 2D. Infection of D17 cells by GP-pseudotyped virus in the absence (lane 1, none) or presence of control (lane 2) or sGP supernatant (lane 3) from transfected 293 cells. Gene transfer was measured by the luciferase assay as described below. Luminescence refers to relative light units in the luciferase assay.

Depend nce of sGP binding n CD16b and correlation of binding with neutrophil activation: Figures 3A-3D.

Neutrophils were incubated for 30 minutes at 4°C with a mouse antibody to CD16b (upper panel; clone 3G8 from Immunotech,

- 10 -

cat# 1M0813) or CD62L (middle panel, R&D Systems), compared to the indicated control antibody [purified mouse IgG (Vector Laboratories), I-2000], followed by supernatants from control or sGP-transfected 293 cells, primary rabbit antibody to sGP, and a FITC-conjugated secondary antibody to rabbit IgG (Fig. 1, legend). Cells were washed with PBS, fixed in 1% formaldehyde, and analyzed by FACS. For blocking, 10^6 cells were incubated with 0.5 – 1 μ g of the relevant antibodies for 30 minutes in a 50 μ l volume.

Figures 3E-3F. Immunostaining with sGP was performed on isolated neutrophils which were maintained in media (none) or incubated with PMA (10 ng/ml) at 37°C for 30 minutes (PMA).

Effect of sGP on neutrophil function: *Figures 4A-4B.* Exposure of neutrophils to sGP inhibits down modulation of L-selectin. Isolated neutrophils were incubated with the indicated control or sGP containing supernatants (Xu, L. et al., *Nat. Med.* (1997) in press) and defined media (AIM V, GIBCO) for 4 hours at 37°C. Expression of L-selectin was determined using an anti-CD62L antibody (R&D Systems), followed by the secondary staining using a FITC-conjugated anti-mouse IgG (Sigma, F2883) as described in Fig. 1, legend. Cells were washed with PBS, fixed with 1% formaldehyde and analyzed by FACS for relative levels of fluorescence intensity as a function of cell number. An isotype control was used to quantitate background levels of immunostaining (neg.). Results are representative of three independent experiments.

II. Results

To determine the specificity of Ebola virus glycoproteins, expression vectors encoding either sGP, GP, or a plasmid control (Xu, L. et al., *Nat. Med.* (1997) in press) were transfected into 293 cells, and cell culture supernatants were used as a source of relevant recombinant glycoproteins. Binding of sGP was determined by immunofluorescence analysis after incubation of relevant supernatants with normal or transformed human cell lines. No binding was detected to several hematopoietic lineages, including lymphocytes or monocytes (Fig. 1A), or transformed Jurkat or CEM T leukemias, the HL60 myelomonocytic or U937 promonocytic leukemia cells. In contrast, sGP was able to bind to granulocytic cells, as evidenced by FACS analysis of this subset of peripheral blood mononuclear cells (PBMC) discriminated by cell size and granularity (Fig. 1A). This cell specificity was confirmed by using double-staining with a granulocyte-specific cell surface marker, CD15 (Fig. 1B). Absorption of sGP

- 11 -

by purified neutrophils in the absence of antibodies also resulted in depletion of sGP, indicating that binding to the neutrophil occurred in the absence of antibody (Fig. 1C).

A potential structural similarity between Ebola GP and avian sarcoma virus envelope protein has been previously proposed (Gallagher, W.R., *Cell* 85:477 (1996)), raising the possibility that this protein could be incorporated into retroviral particles.

To determine the binding specificity of the transmembrane glycoprotein, pseudotyping of a Moloney leukemia virus was therefore attempted. Infectivity of different cell types by this pseudotyped vector was determined with a luciferase reporter gene. de Wet, J.R. et al., *Mol. Cell. Biol.* 7:725 (1987). This analysis revealed infection of cells

different from those which interacted with sGP (Fig. 2A,B). For example, though it could infect other cell types, transduction by the GP retroviral vector readily occurred in endothelial cells, either from the microvasculature (MVEC) or umbilical veins (HUVEC) (Fig. 2B), which did not bind sGP (Fig. 2C, left). When the specificity of GP-retrovirus was compared to murine retroviruses pseudotyped with amphotropic or ecotropic envelope gp70 proteins, the range of susceptible target cells differed (Fig. 2B), suggesting that the virus receptor(s) for Ebola GP differ from those previously described for gp70. Minimal binding of GP-virus was observed on neutrophils, despite the ability of these cells to bind sGP (Fig. 2C, upper panel) and the fact that immunoreactive protein was detected on the virus. Conversely, GP-virus binding to endothelial cells was readily detected, though these cells did not bind sGP (Fig. 2C, lower panel). When sGP was analyzed for its effect on GP retroviral gene transfer, infection was not inhibited by sGP (Fig. 2D), further confirming the divergent specificities of the two forms of the viral glycoprotein. Recent studies have revealed that the biochemical forms of these proteins differ, with sGP present in solution primarily as a homodimer and GP as a trimer, suggesting that differences in multimer composition may contribute to these alternative specificities.

Potential cell surface receptors for sGP were analyzed with antibodies to several neutrophil cell surface antigens to interfere with sGP binding, including CD15, L-selectin (CD62L), CD16b, and several common leukocyte antigens. Only the neutrophil-specific form of the low affinity $F_c \gamma$ receptor III, CD16b, inhibited sGP binding specifically. Antibodies to CD62L, for example, did not inhibit sGP binding (Fig. 3). Binding to neutrophils correlated with their activation state and CD16b expression since no binding was observed in cells stimulated with phorbol 12-myristate 13-acetate (PMA) for 30 minutes, at which time CD16b expression was markedly decreased on these cells (Fig. 3, lower panel). Overexpression of this form

- 12 -

of CD16 on a heterologous cell type, 3T3 fibroblasts, did not confer sGP binding to these cells by FACS analysis, suggesting that CD16b is necessary but not sufficient for stable binding.

Binding of sGP did not inhibit neutrophil activation in response to potent pleiotropic activators (PMA, IL-8, or f-Met-Leu-Phe), as measured by down modulation of L-selectin expression using FACS analysis. In a defined serum-free medium, partial activation of neutrophils was observed, with a decrease in L-selectin expression at 4 hours (Fig. 4). Under these conditions, incubation of neutrophils with sGP supernatant prevented this decrease in L-selectin expression (Fig. 4). Because L-selectin was not required for sGP binding (Fig. 3), this effect was apparently indirect, through a mechanism not yet defined, possibly involving CD16b or carbohydrate interactions of the highly glycosylated sGP protein.

The expression of alternative Ebola virus glycoproteins in clinical infection has long been recognized, but their functional roles and cell specificity have not been defined. Early after infection, high levels of the secreted protein are found in the serum and precede fulminant replication and dissemination of virus systemically, at which time synthesis of transmembrane GP is markedly increased. Sanchez, A. et al., *PNAS (USA)* 93:3602 (1996). The inventors have now found that the binding specificities of these two molecules differ. It had been proposed that sGP may serve as a decoy to prevent recognition of GP, possibly to temporarily inhibit virus binding to target cells. The studies set forth herein suggest that this hypothesis is unlikely to be correct. The binding specificities of these proteins differ, and despite the fact that they are derived from the same viral gene, it has been surprisingly found that alternative forms of the glycoprotein have been selected for different functions.

Although these proteins share identical amino terminal sequences, their carboxyl terminal regions differ. Sanchez, A. et al., *Virus Res.* 29:215 (1993). These sequences are likely responsible for the differences in binding specificity, either through direct interactions in these domains or by their effect on multimerization. The secreted glycoprotein binds to neutrophils to prevent early events in activation, possibly serving to diminish any inflammatory responses which might provide innate immunity to the virus, facilitating productive viral replication. The subsequent increase in GP synthesis gives rise to virus which in turn could infect other cells. Filoviruses have been shown previously to infect and replicate in different cell types and appear to grow readily in endothelial cells *in vivo*. Peters, C.J. et al., *Fields Virology*, B.N. Fields, D.M. Knipe and P.M. Howley, Eds. (Lippincott-Raven, Philadelphia) (1996);

Schnittler, H.J. et al., *J. Clin. Invest.* 91:1301 (1993). The findings set forth herein suggest that its tropism for this cell type is probably determined by the specificity of GP. In Ebola infection, preferential binding and infection of microvascular endothelial cells may lead ultimately to a loss of capillary integrity that results in the severe hemorrhage observed in the terminal stages of this disease. The differential binding of these two gene products from the same viral structural gene generated by RNA editing suggests that they have evolved functionally to differentially affect immunity and infectivity. The ability to facilitate viral replication and target the virus to endothelial cells by alternative products of the same viral gene represents an efficient genetic mechanism which can account for different pathologic features of this disease. Inhibition of sGP binding to neutrophils and GP to endothelium is likely to ameliorate the effects of acute Ebola virus infection.

SPECIFIC EXAMPLE 2

I. Methods

Production of pseudotyped MuLV retroviruses expressing green fluorescent protein (GFP): 50% - 70% confluent 293 T cells in 60mm tissue culture dishes were transfected using the calcium phosphate method and the following plasmids: 0.3 μ g 1012 GP(Z) (see Figure 8) or 1012 sGp-Gp(Z) (see Figure 9), 3 μ g LZR-gfp, 2 μ g pNGVL-gag-pol. After overnight transfection, fresh media was added to cells. Twenty hours later, the supernatants were harvested and filtered through a .45 μ m filter.

Infection of HUVEC cells using the pseudotyped retroviruses: The day before infection, 30% - 50% confluent HUVEC cells were prepared in 6-well plates. 1 ml of pseudotyped retroviral supernatant was added to one well of the 6-well plates with 15 μ g/ml of polybrene. Sixteen hours later, the viruses were removed and normal media was added. After 24 hours, the cells were lifted and GFP expression measured using FACS analysis.

Construction of 1012 sGP-GP(Z): 1012 sGP(Z) cells were digested with PstI and treated with Klenow, then digested with XbaI. 1012 GP(Z) cells were digested by EcoRI and treated with Klenow, then digested with KpnI. PstI/Klenow/XbaI treated sGP fragment and EcoRI/Klenow/KpnI treated GP fragment were then cloned into XbaI/KpnI treated pVR-1012 plasmid.

GP and sGP derivatives: The receptor recognition domain, mucin-like domain and/or TM domain of GP and sGP were mutated. The mutated GP and sGP was then tested for its ability to pseudotype and for cytotoxicity in producer cells.

- 14 -

II. Results

To determine the efficacy of targeting endothelium with the gene transfer vectors pseudotyped with GP of the present invention, HUVEC cells were infected with GP(Z) pseudotyped MuLV retrovirus (LZR-gfp) and sGP-GP(Z) pseudotyped MuLV retrovirus (LZR-gfp). Figures 5A-5C show the infection rate (GFP expression) measured using FACS analysis. As shown in Figure 5B, the GP(Z) pseudotyped MuLV retrovirus (LZR-gfp) was effective in targeting and expressing GFP in endothelium.

To determine whether mutating GP would effect its ability to pseudotype and/or decrease toxicity in producer cells, the receptor recognition domain, mucin-like domain and/or TM domain were mutated. Figure 10 shows the results. The optimal envelope is able to pseudotype but shows minimal toxicity.

The foregoing discussion discloses and describes merely exemplary embodiments of the present invention. One skilled in the art will readily recognize from such discussion, and from the accompanying drawings and claims, that various changes, modifications and variations can be made therein without departing from the spirit and scope of the invention as defined in the following claims.

All patents and other references cited herein are incorporated by reference as if fully set forth.

- 15 -

WE CLAIM:

1. A genetic construct comprising a gene operatively-linked to a carrier, wherein the carrier is associated with a transmembrane form of viral glycoprotein or derivative thereof.
- 5 2. The genetic construct of Claim 1, wherein the transmembrane form of viral glycoprotein or derivative thereof is expressed on the surface of the carrier.
3. The genetic construct of Claim 1, wherein the transmembrane form of viral glycoprotein or derivative thereof is from Ebola.
4. The genetic construct of Claim 1, wherein the carrier is a viral vector.
- 10 5. The genetic construct of Claim 1, wherein the carrier is a non-biologic gene targeting vehicle.
6. The genetic construct of Claim 4, wherein the viral vector is a retroviral vector.
7. The genetic construct of Claim 4, wherein the viral vector is a lentiviral
15 vector.
8. The genetic construct of Claim 5, wherein the non-biologic gene targeting vehicle is a liposome.
9. The genetic construct of Claim 5, wherein the non-biologic gene targeting vehicle is a DNA-protein complex.
- 20 10. A method of targeting a gene to a cell comprising the step of administering to a cell population a genetic construct comprising the gene operatively-linked to a carrier, wherein the carrier is associated with a transmembrane form of viral glycoprotein or derivatives thereof.
- 25 11. The method of Claim 10, wherein the transmembrane form of viral glycoprotein or derivative thereof is expressed on the surface of the carrier.

- 16 -

12. The method of Claim 10, wherein the transmembrane form of viral glycoprotein or derivative thereof is from Ebola.
- 13. The method of Claim 10, wherein the carrier is a viral vector.
- 14. The method of Claim 10, wherein the step of administration is *ex vivo*.
- 5 15. The method of Claim 10, wherein the step of administration is *in vivo*.
16. The method of Claim 10, wherein the cell is an endothelial cell.
17. The method of Claim 10, wherein the cell is a hepatocyte.
18. The method of Claim 10, wherein the cell is a monocyte.
19. The method of Claim 10, wherein the cell is a dendritic cell.
- 10 20. The method of Claim 14, further comprising the step of introducing the cell population to a subject.

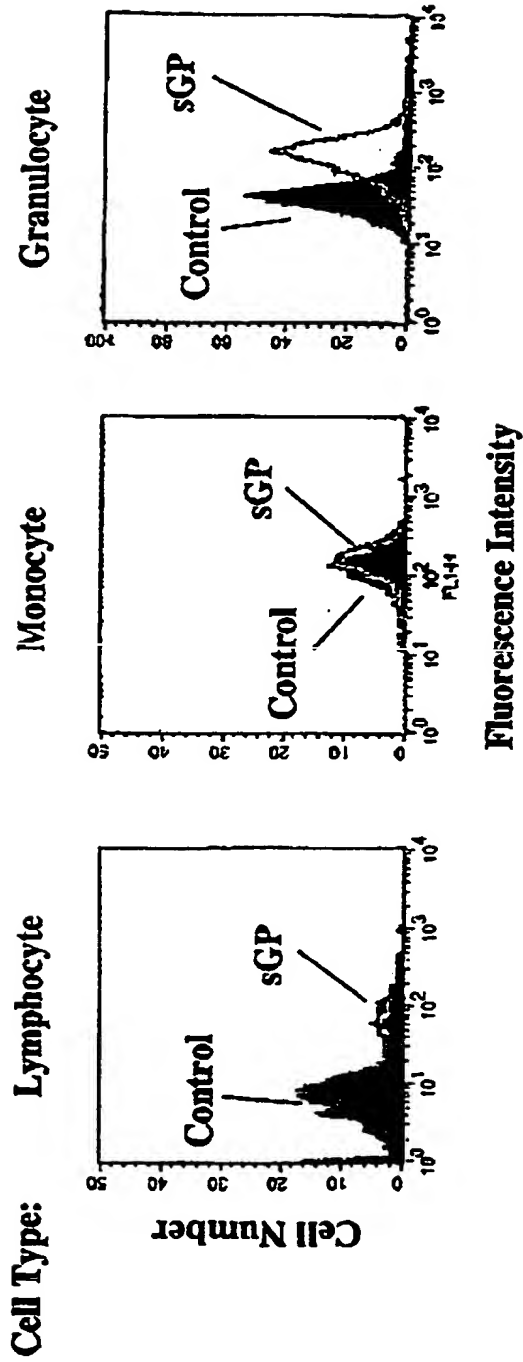


Figure 1A

Figure 1A1

Figure 1A2

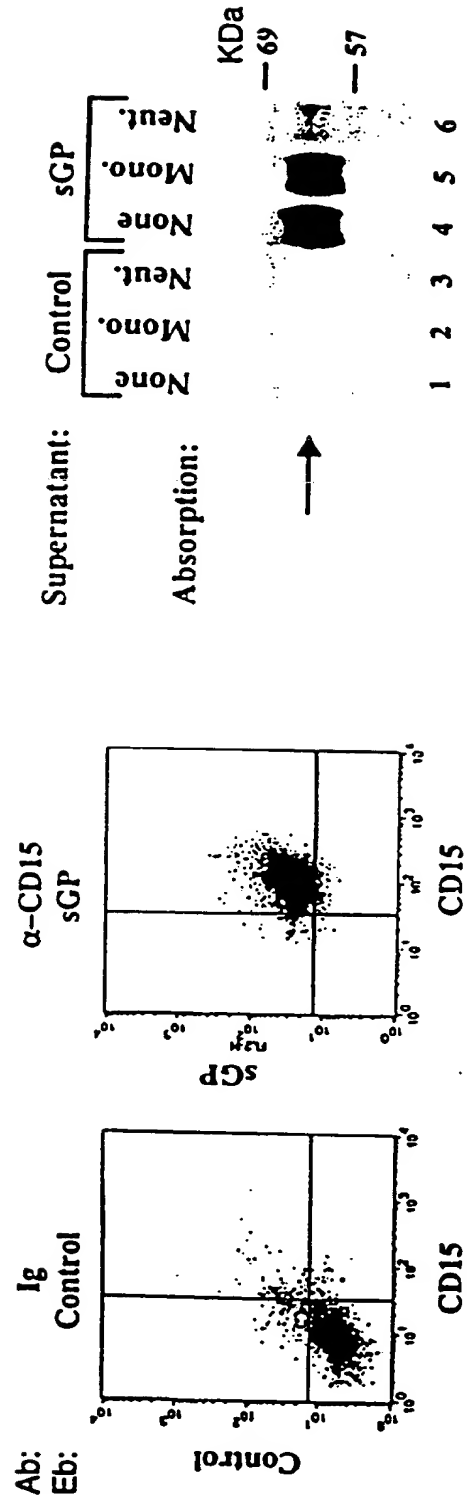


Figure 1B

Figure 1B1

Figure 1C

2/11

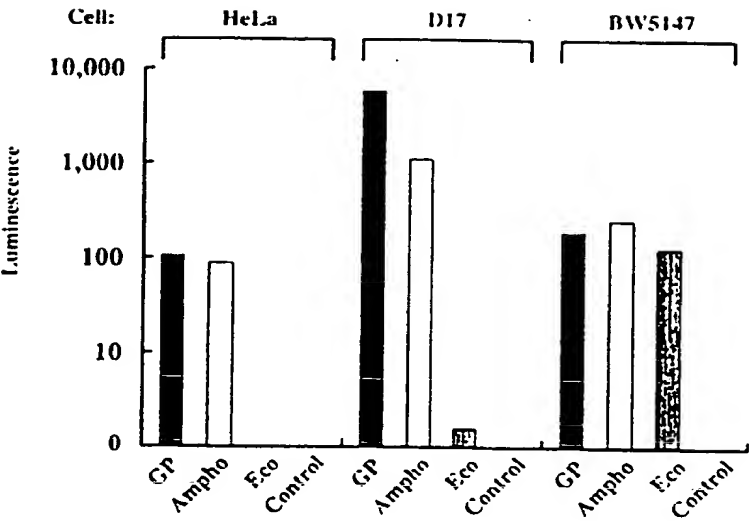


Figure 2A

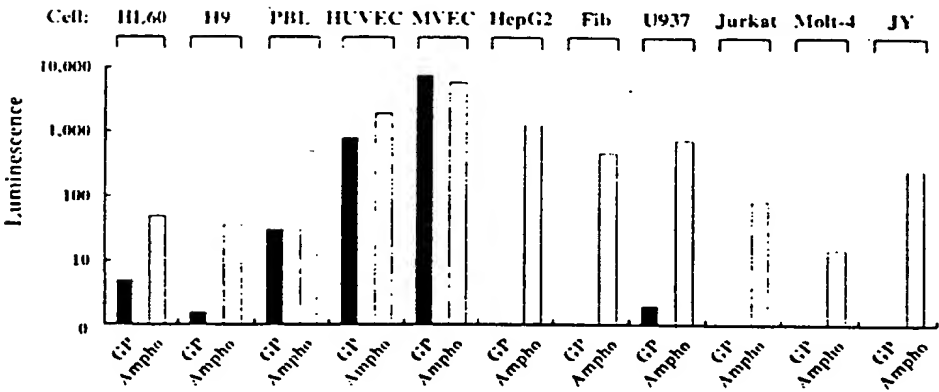
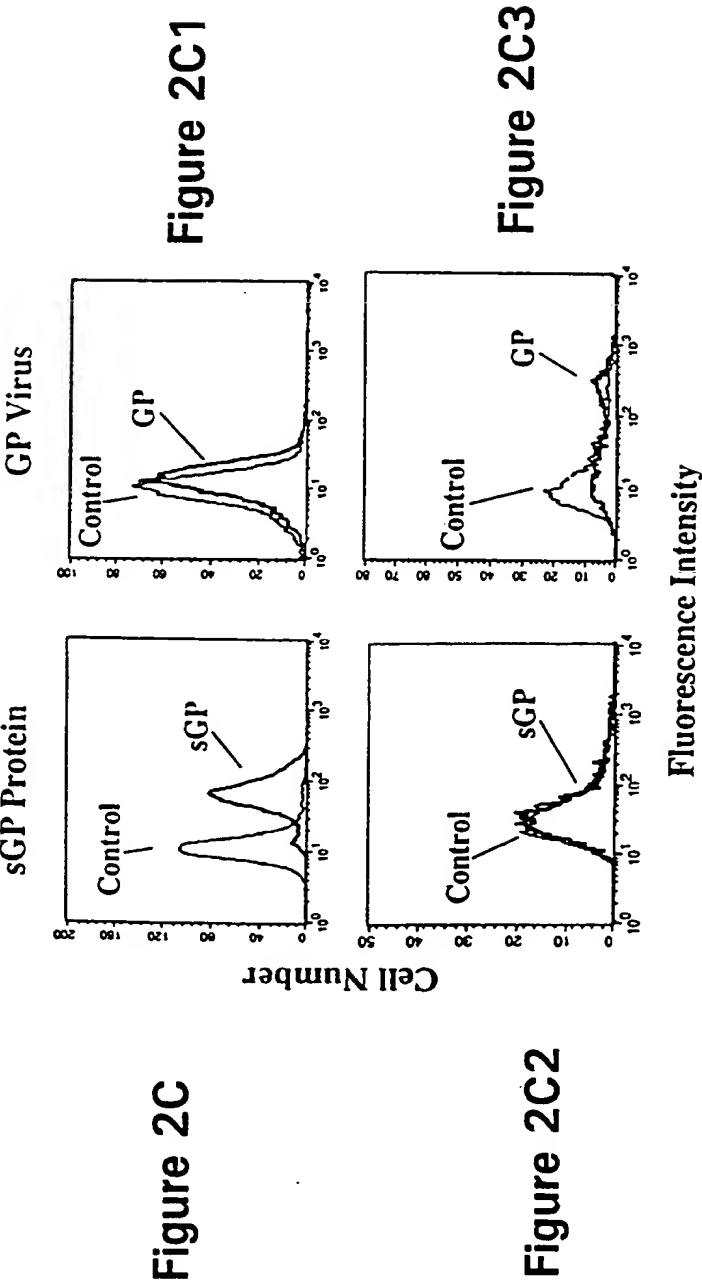


Figure 2B



4/11

Figure 3A

Ab: Control (Ig)

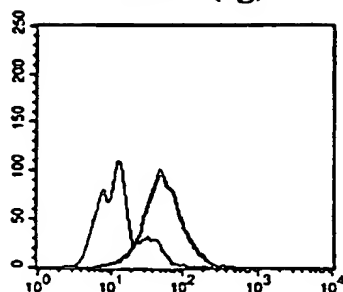


Figure 3B

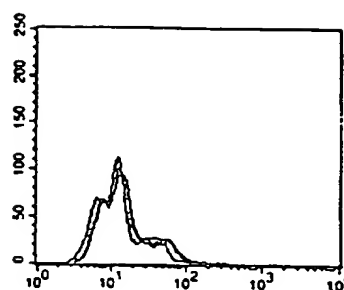
 α -CD16

Figure 3C

Ab: Control (Ig)

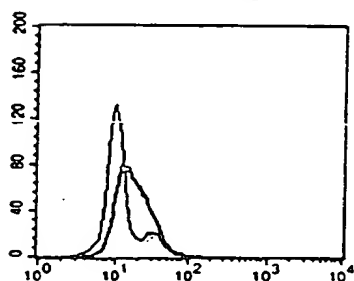
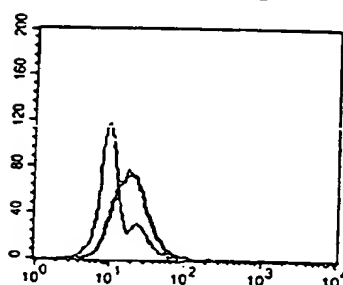


Figure 3D

 α -CD62L

Cell Number

Figure 3E

Stim: None

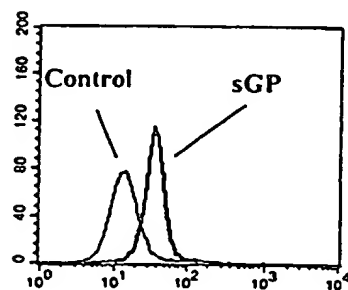
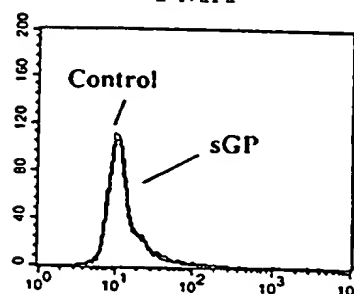


Figure 3F

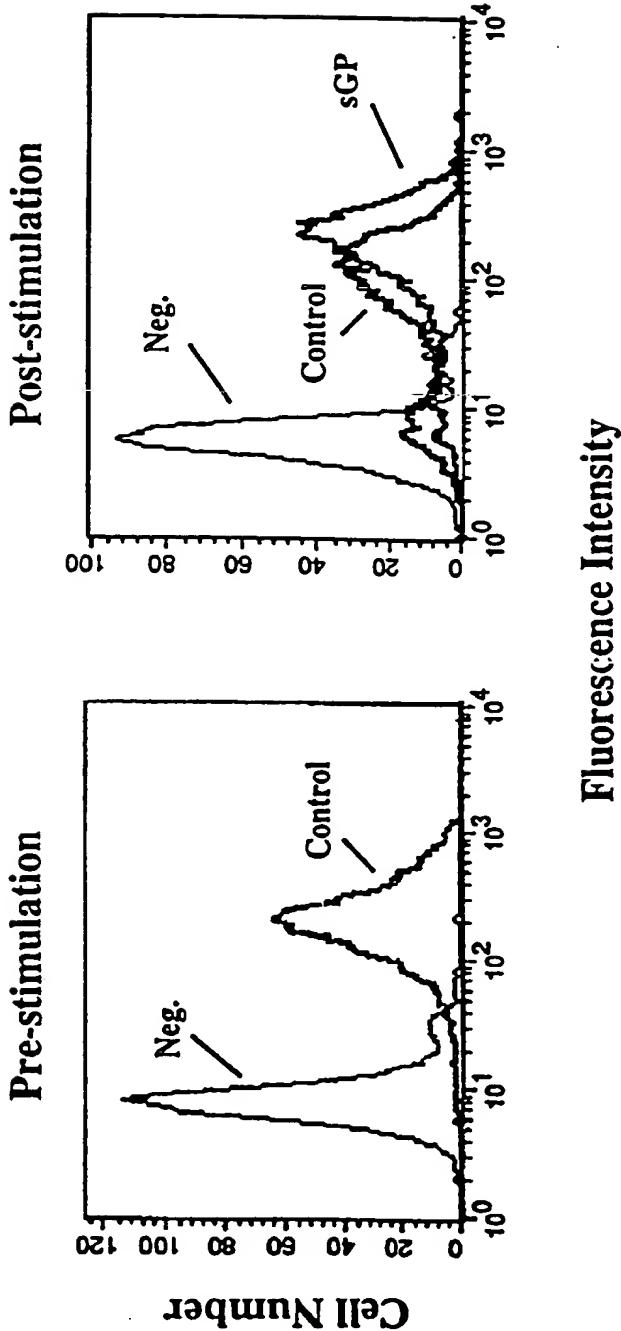
PMA



Fluorescence Intensity

5/11

Figure 4A



6/11

Figure 5A

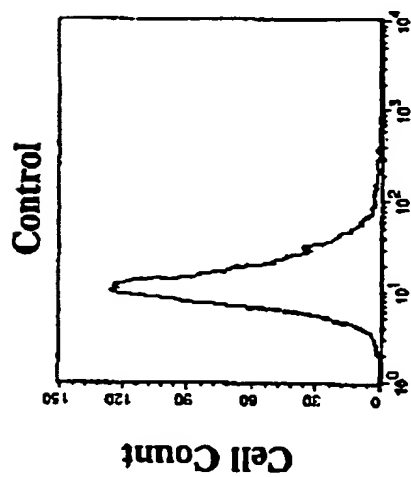


Figure 5B

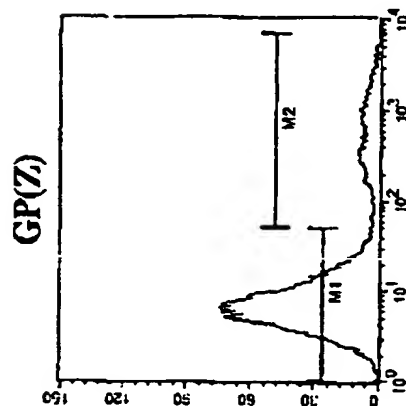
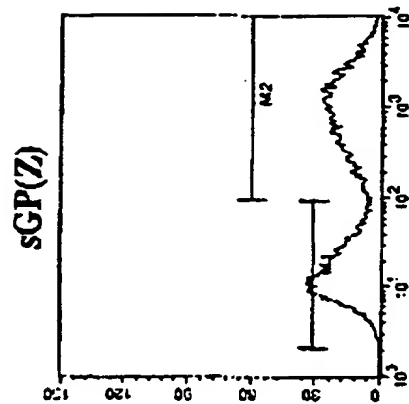
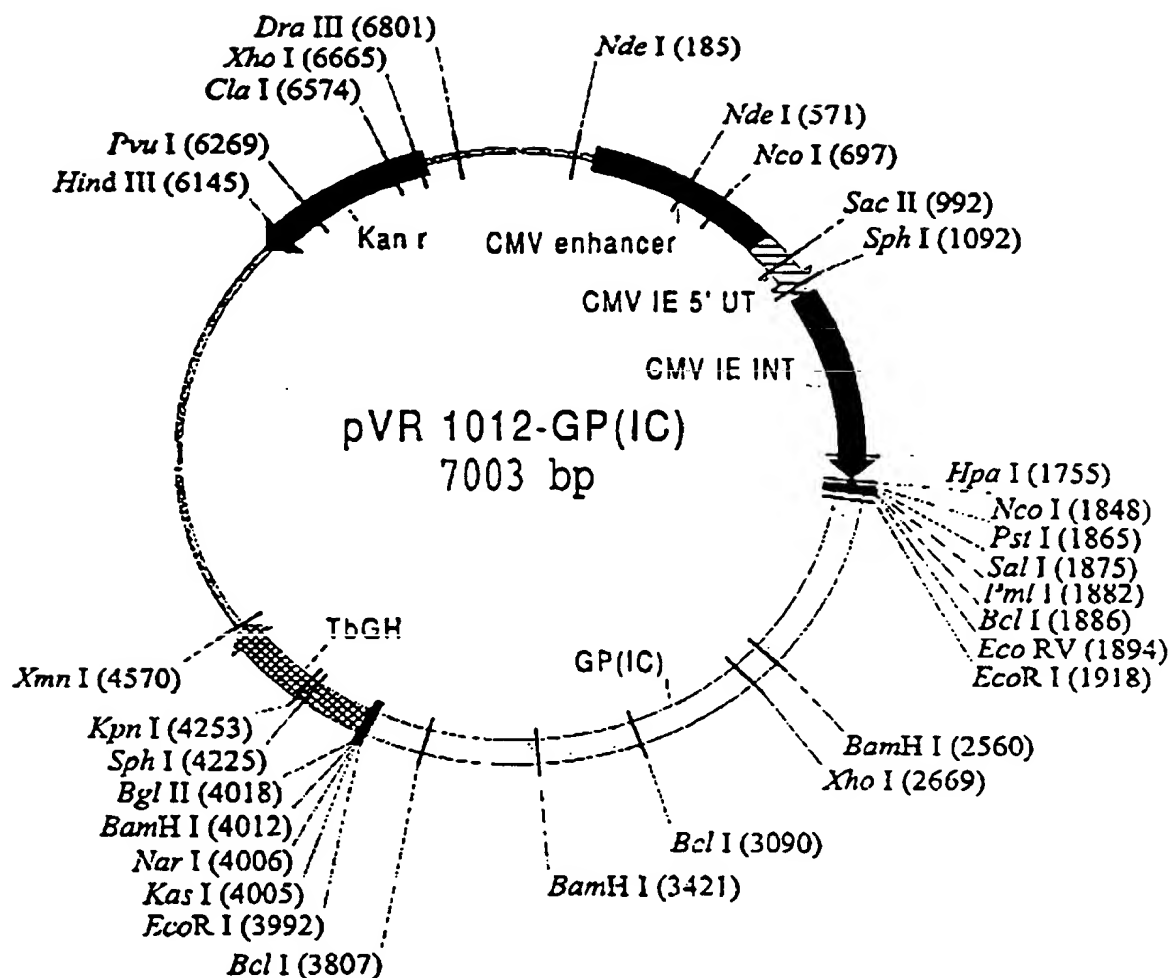


Figure 5C



Fluorescence Intensity

7/11

**Figure 6**

8/11

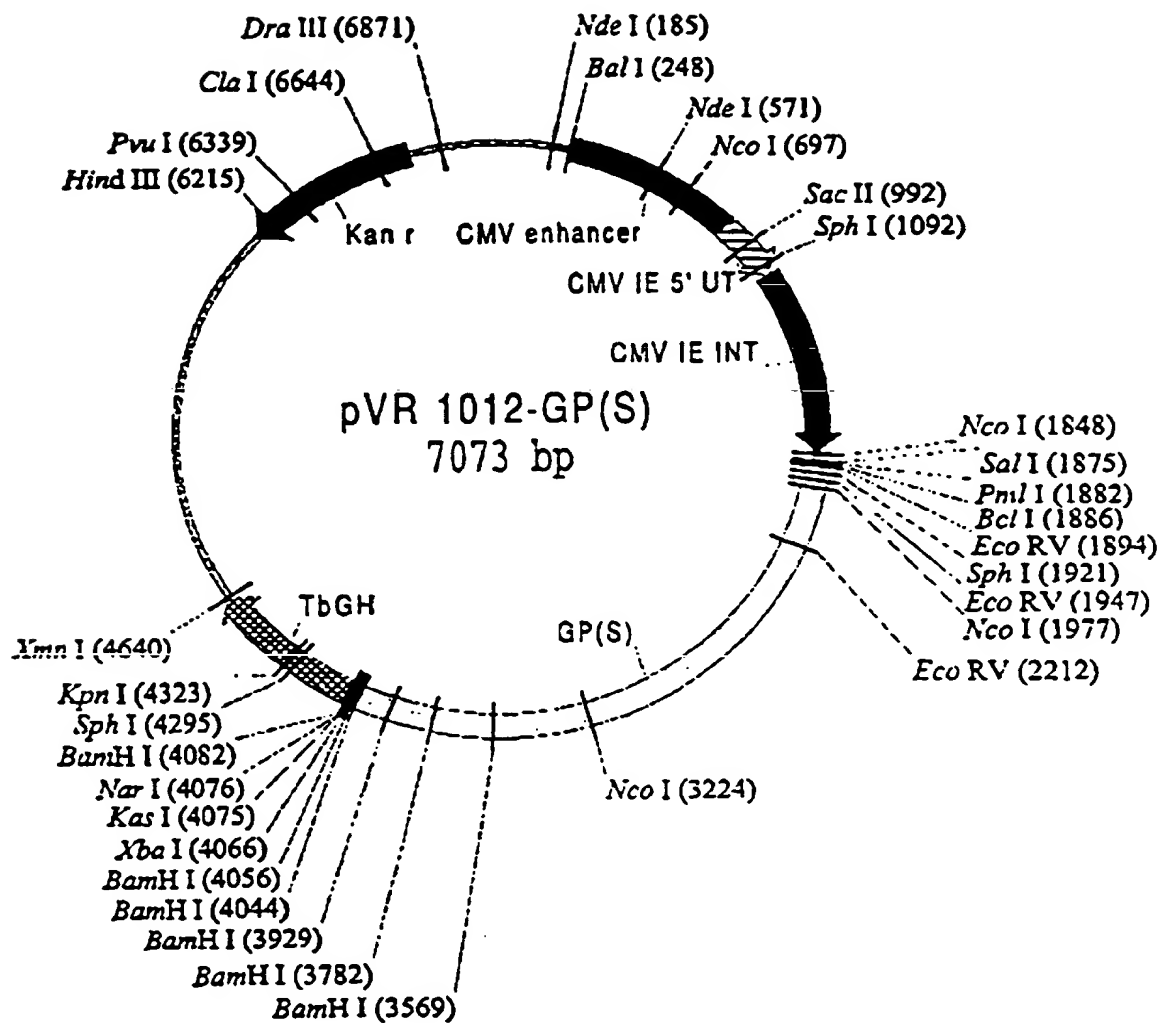


Figure 7

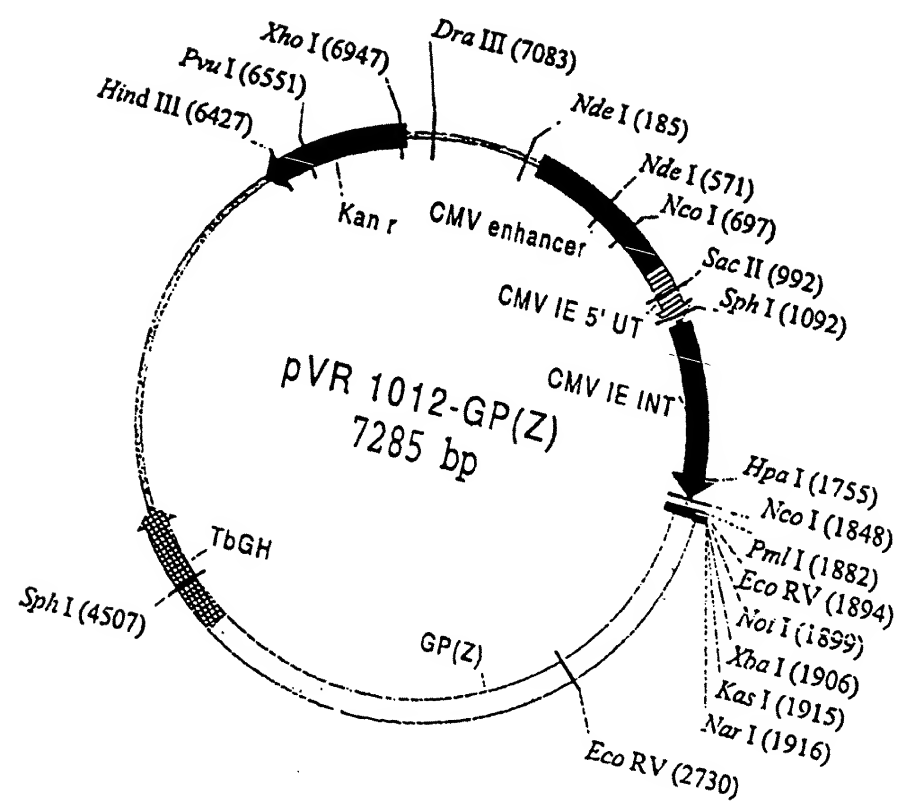


Figure 8

10/11

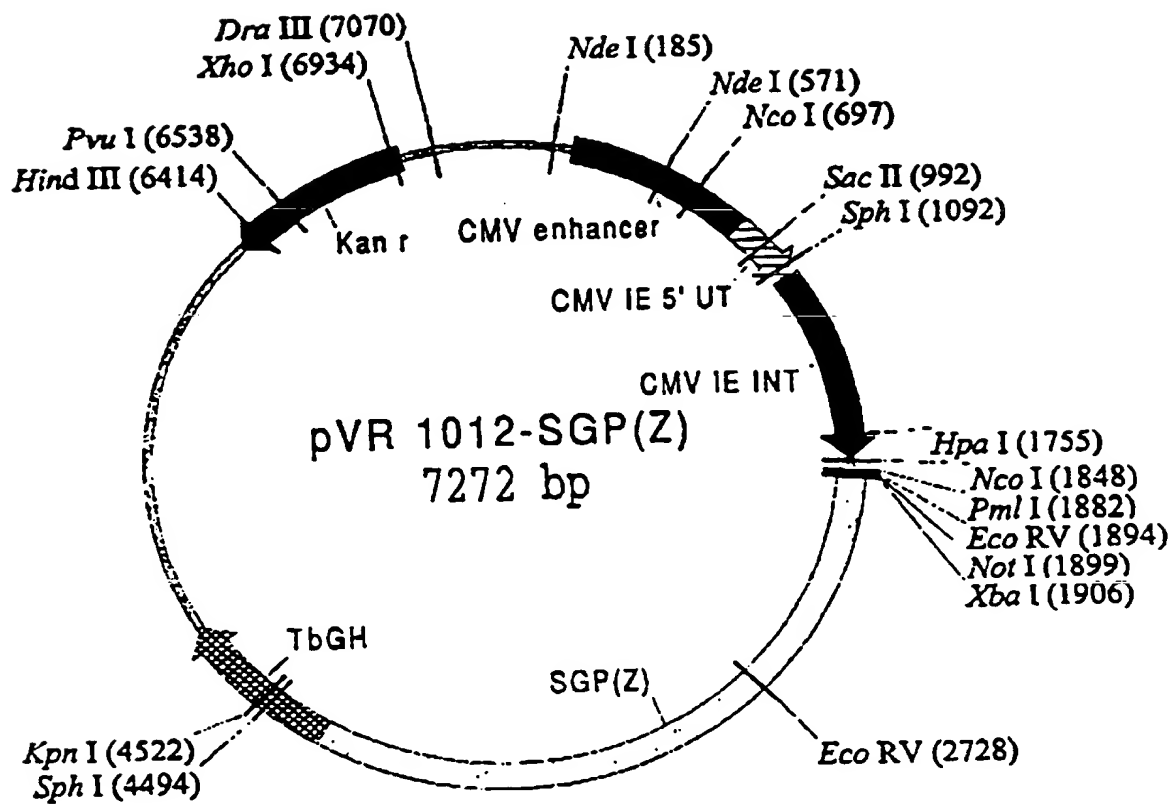


Figure 9

11/11

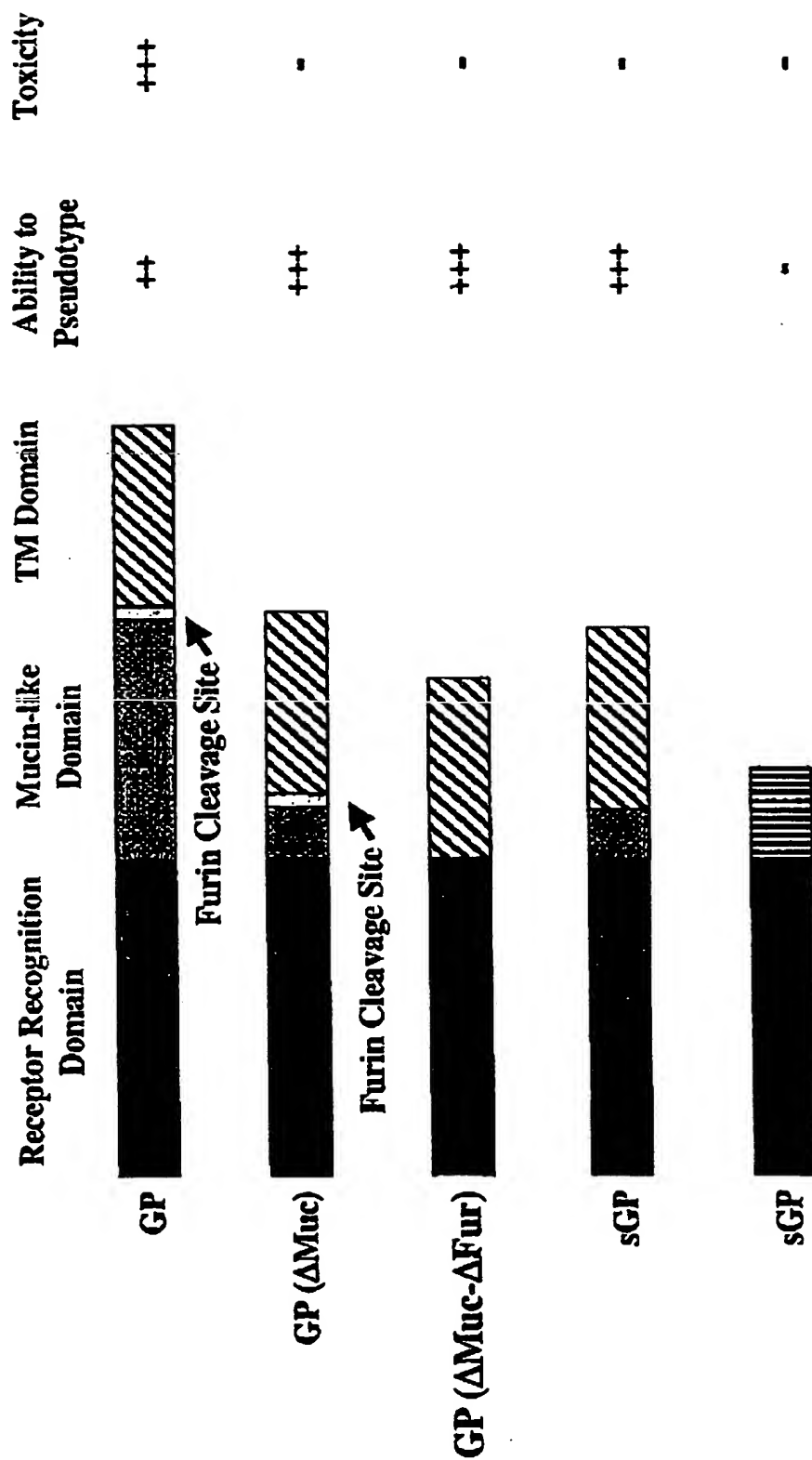


Figure 10

SEQUENCE LISTING ID NO: 1

pVR 1012-GP(IC)

General Description

DNA pVR 1012-GP(IC)
Local object
Created: 09/14/98 04:17PM
Last Modification Date: ? (no data)
length: 7003 bp
storage type: Basic
form: Circular

Comments

Restriction Map

BglII: 1 site AGATCT
 TCTAGA

Clal: 1 site ATTCAT
 TAGCTA

DraIII: 1 site CACNNNGTC
 GTGNNNCAC

EcoRV: 1 site GATATC
 CTATAG

HindIII: 1 site AAGCTT
 TTCCAA

HpaI: 1 site GTTAAC
 CAATTG

KasI: 1 site GGCGCC
 CCGCGG

KpnI: 1 site GGTACC
 CCATGG

NarI: 1 site GGCGCC
 CCGCGG

PmlI: 1 site CACGTG
 GTGCAC

PstI: 1 site CTGCAG
 GACCTC

PvuI: 1 site CGATCG
 GCTAGC

SacII: 1 site CCGCGG
 GGCGCC

SalI: 1 site GTCGAC
 CAGCTG

XmnI: 1 site GAANNKNTTC
 CTTNNNNAAG

EcoRI: 2 sites GAATTC
 CTTAAG

NcoI: 2 sites CCATGG
 GGTACC

NdeI: 2 sites CATATG
 GTATAC

SphI: 2 sites GCATGC
 CGTACG

XhoI: 2 sites CTCGAG
 GACCTC

BamHI: 3 sites GGATCC
 CCTAGG

BclI: 3 sites TGATCA
 ACTAGT

Functional Map

CDS (4 signals)

CMV IE 5' UT

Start: 886 End: 1129

CMV IE INT

Start: 1130 End: 1840

TbGH

Start: 4020 End: 4572

Kan^r

Start: 6068 End: 6690 (Complementary)

Misc_feature (2 signals)

CMV enhancer

Start: 248 End: 885

GP(IC)

Start: 1870 End: 4019

Annotations

```

1  TCGCGCGTTT CGGTGATGAC GGTGAAAACC TGTGACACAT GCAGCTCCCG
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NdeI

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.....
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NdeI

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NcoI

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NcoI

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SacII

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SphI

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1201 TATTGGTGAC GATACCTTCC ATTACTAATC CATAACATGG CTCTTTGSCA
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1251 CAACATATCT TATTGGCTAT ATGCCAATAC TCTGTCTTTC AGAGACTGAC
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1701 CTGAGTTGTT GTATTCTGAT AAGAGTCAGA GGTAACTCCC GTTGCGGTGC
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EpaI

1751 TGTAAACGGT GGAGCCGAGT GTACTCTGAG CAGTACTCGT TGCTGCCCGG
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NcoI

1801 CGCGCCACCA GACATAATAG CTGACAGACT AACAGACTGT TCCTTTCCAT
 CGCGCGTGGT CTGTATTATC GACTGTCTGA TTGTCTGACA AGGAAAGCTA

SaIINcoIPstIPmlIBclIEcoRV

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EcoRI

1901 CGCGCGGGGC GCTCTAGAAT TCTCTAATCA CAGTCATCAT GCGAGCGTCA
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2001 TTGGGTAATA ATCCTATTCC ATAAAGTCTT TTCAATCCCG TTGGGGGTTG
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2051 TACACAACAA TACCCTACAA GTGAGTGATA TCGACAAGTT TGTGTGCCGA
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2101 GACAAACTCT CTTCAACTAG CCAATTGAAG TCAGTCGGGT TGAACCTGGA
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2151 GGGCAATGGA GTAGCAACTG ATGTACCAAC GCGAACCAAA AGATCGGGTT
 CCCGTTACCT CATCGTTGAC TACATGGTTC CCGTTGGTTT TCTACCCCAA

2201 TTCGAGCTGC TGTTCACCA AAGGTGGTAA ATTACGAAC TGGAGAATGG
 AAGCTCGACC ACAAGGTGGT TTCCACCATT TAATGCTTCG ACCTCTTACC

2251 GCTCAGAACT GTTATAACCT GGCTATAAAG AAAGTTGATG GTAGTGAGTC
 CGACTCTTGA CAATATTGGA CCGATATTTC TTTCAACTAC CATCACTCAC

2301 CCTACCAGAA GCGCCTGAGG CAGTGAGGGA TTTCCCGGTT TGCCGCTATG
 GGATGGTCTT CGGGGACTCC CTCACTCCCT AAAAGGGGCA ACGGCGATAC

2351 TACACAAAGT CTCAGGAAGT GGACCATGCC CAGGAGGACT CGCCTTTCAC
 ATGTGTTTCA GAGTCCTTGA CCTGGTACGG GTCCTCCTGA CCGGAAAGTG

2401 AAAGAAGCAG CCTTCTTCCT GTATGACCGA CTCGCATCAA CAATCATTTA
 TTTCTTCCTC GGAAGAAGGA CATACTGGCT GAGCGTAGTT GTTAGTAAT

2451 TCGGGGTACA ACCTTTCCCG AAGGAGTTAT TGCAATTTCTG ATCTTGCCTA
 AGCCCCATGT TCGAAACGGC TTCTTCAATA ACGTAAAGAC TAGAACGGAT

2501 AGGCGCGAAA GGATTTTTTC CAGTCTCCTC CATTCATGA GCTGCCAAC
 TCCGCGCTTT CCTAAAAAG GTCAGAGGAG GTAACGTACT CGGACGGTTC

BamHI

2551 ATGACCACGG ATCCCTCCAG TTACTATCAC ACGACAACAA TAAACTACGT
TACTCGTGCC TAGGGAGGTC AATGATAGTG TGCTGTTGTT ATTTGATGCA

2601 GGTTCATAAT TTTGGAACCA ACACCACAGA GTTTCGTGTC CAAGTCGATC
CCAACATATTA AAACCTTGGT TGTGGTGTCT CAAAGACAAAG GTTCAGCTAG

XhoI

2651 ATTTGACGTA TOTGCAGCTC GAGGCAAGAT TCACACCACA ATTCTTGTG
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2701 CTCCTAAATG AAACCATCTA CTCTGATAAC CGCAGAAGTA ACACAACAGG
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2751 AAAACTAATC TGGAAAATAA ATCCCACTGT TGATACCAGC ATGGGTGAGT
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2801 GGGCTTCTG GGAATAATAA AAAACTTCAC AAAAACCTT TCAAGTGAAG
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2851 AGTTGTCTTT CGTACCTGTA CCAGAAACCC AGAACCCAGT CTTGACACG
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2901 ACAGCGACGG TCTCTCTCC CATCTCCGCC CACAACCAGC CAGGCGAAGA
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2951 CCACAAAGAA TTGGTTTCAG AGGATTCCAC TCCAGTGGT CAGATGCAAA
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3001 ACATCAACGG AAGGACACA ATGCCAACCA CAGTCACGGG TGTAACCAACA
TGTACTTCCC TTTCTGTGT TAGGGTGGT GTCACCTCCC ACTGCTTGT

BclI

3051 ACCACACCTT CTCCATTTCC AATCAATGCT CGCAACACTG ATCATACCAA
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3101 ATCATTATC GGCCTGGAGG GCCCCAAGA AGACCACAGC ACCACACAGC
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3151 CTGCCAAGAC CACCAGCCAA CCAACCAACA GCACAGAATC GACGACACTA
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3201 ATCCCAACAT CAGAGCCCTC CAGTAGAGGC ACGGACCAT CCAGCCCCAC
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3251 GGTCCCCAAC ACCACAGAAA CCCACGCCGA ACTTGGCAAG ACAACCCCA
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3351 CACCCCGACG AACTCAGTGG ACCTGGCTTC CTGACGAACA CAATACGGGG
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BamHI

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BclI

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EcoRI

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NarIXasIBamHI BglII

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SphI

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KpnI

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4501 GCTATTAAAT GCAGAGGGAG AGAAATGCC TCCAACATGT GAGCAAGTAA
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XbaI

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5001 CTGTCTCCAC GAACCCCCCG TTCAGCCCGA CCGCTGCGCC TTATCCGGTA
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6051 ATCAATACAA CCTATTAAAT TCCCTCCTC AAAAATAAGG TTATCAAGTC
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HindIII

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6151 TGCATTTCTT TCCAGACTTG TTCAACAGGC CAGCCATTAC GCTCGTCATC
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6201 AAAATCACTC GCATCAACCA AACCGTTATT CATTCGTGAT TGCCTCAG
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PvuII

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6301 GAATGCAACC GCGCGAGSAA CACTGCCAGC GCATCAACAA TATTTTCACC
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6402 CAGTGGTGAG TAACCATGCA TCATCAGGAG TACGGATAAA ATGCTTGATG
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6451 GTCGGAAGAG GCATAAATTC CGTCAGCCAG TTTAGTCTGA CCATCTCATC
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6501 TGTAACATCA TTGGCAACCG TACCTTTGCC ATGTTTCAGA AACAACTCTG
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ClaI

6551 GCGCATCGGG CTTCCCATAC AATCGATAGA TTGTCCACC TGATTGCCCC
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6601 ACATTATCGC GAGCCCATTT ATACCCATAT AATCAGCAT CCATGTTGGA
TGTAATAGCG CTCGGSTAAA TATCGGTATA TTTAGTCGTA GGTACAACT

XhoI

6651 ATTTAATCGC GCGCTCGAGC AAGACGTTTC CCGTTGAATA TGGCTCATAA
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6701 CAGCCCTTGT ATTACTGTTT ATGTAAGCAG ACAGTTTTAT TGTTTCATGAT
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DraIII

6751 GATATATTTT TATCTTGTC AATGTAACAT CAGAGATTTT GAGACACAC
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DraIII

6801 GTGGCTTTCC CCCCCCCCCC ATTATTGAAG CATTATCAG GGTATTGTC
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6851 TCATGAGCCG ATACATATTT GAATGTATTT AGAAAAATAA ACAAAATAGGG
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6901 GTTCCGCCCA CATTTCCTCCG AAAAGTCCCA CCTGACGTCT AACAAACCAT
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6951 TATTATCATG ACATTACCT ATAAAAATAG GCGTATCAGG ACGCCCTTTC
ATAATAGTAC TCTAATTGGA TATTTTATC CCCATAGTGC TCCGGGAAG

7001 CTC
CAG

pVR 1012-GP(S)

General Description

DNA pVR 1012-GP(S)

Local object

Created: 09/14/98 03:58PM

Last Modification Date: ? (no data)

length: 7073 bp

storage type: Basic

form: Circular

Comments

Restriction Map

BamI: 1 site TGGCCA
ACCGGTBclI: 1 site TCATCA
ACTAGTClaI: 1 site ATCGAT
TAGCTADraIII: 1 site CACGNGTG
CTGNNKACHindIII: 1 site AAGCTT
TTCGAAKasI: 1 site GCGGCC
CCGCGGKpnI: 1 site GGTACC
CCATGGNarI: 1 site GCGGCC
CCGCGGPmlI: 1 site CACGTG
GTGCACPvuI: 1 site GGATCG
GCTAGCSacI: 1 site CCGCGG
GGCGCCSall: 1 site GTCCAC
CAGCTGXbaI: 1 site TCTAGA
AGATCTXmnI: 1 site GAANNNTTC
CTTNNNAAGNdeI: 2 sites CATATG
GTATACEcoRV: 3 sites GATATC
CTATAGSphI: 3 sites GCATGC
CGTACGNcoI: 4 sites CCATGG
GGTACCBamHI: 6 sites GCATCC
CCTAGG

Functional Map

CDS (4 signals)

CMV IE 5' UT

Start: 886 End: 1129

CMV IE INT

Start: 1130 End: 1840

TbGH

Start: 4090 End: 4642

Kan r

Start: 6138 End: 6760 (Complementary)

Misc_feature (2 signals)**CMV enhancer**

Start: 248 End: 885

GP(S)

Start: 1870 End: 4089

Annotations

1 TCCCGCGTTT CCGTGATGAC GGTGAAAACC TCTGACACAT GCAGCTCCCC
 ASCGCGCAAA GCCACTACTG CCACTTTTGG AGACTGTGTA CGTCGAGGGC

 51 GAGACGGTCA CAGCTTGTCT GTAAAGCGGAT GCCGGGAGCA GACAAGCCCC
 CCTCGCACT GTCGAACAGA CATTGCGCTA CGGCCCTCGT CTGTTGCGGC

 101 TCAGGGCCCG TCAGCGGGTG TTGCGGGGTG TCGGGGCTGG CTTAACTATG
 AGTCCGCGC AGTCGCCCCA AACCGCCCCA AGCCCCGACC GAATTGATAC

NdeI

151 CGGCATCAGA CCAGATTGTA CTGAGAGTGC ACCATATGCG GTGTGAATA
 CGCGTAGTCT CGTCTAACAT GACTCTCAGG TGGTATACGC CACACTTTAT

BalI

201 CCGCACAGAT CGGTAAGGAG AAAATACCGC ATCAGATTGG CTATTGGCCA
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 251 TTGCATACGT TGTATCCATA TCATAATATG TACATTATA TTGGCTCATG
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 351 AATCAATTAC GGGGTCATTA GTTCATAGCC CATATATGGA GTTCCGCGTT
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 451 CCCATTGACG TCAATTAATGA CGTATGTTCC CATAGTAACG CCAATAGGGA
 GGGTAACGCG AGTTATTACT GCATACAAGG GTATCATTGC GGTATCCCT

 501 CTTTCCATTG ACGTCAATGG GTGAGTATT TACGGTAAAC TGCCCACTTG
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NdeI

551 GCACTACATC AAGTGTATCA TATGCCAAGT ACGCCCCCTA TTGACGTCAA
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 601 TGACGGTAAA TGGCCCCCCT GGCATTATGC CCAGTACATG ACCTTATCGG
 ACTGCCATTI ACCGGGCGGA CCGTAATACG GGTCACTGAC TGAATACCC

NcoI

651 ACTTCCTAC TTGGCAGTAC ATCTACGTAT TAGTCATCGC TATTACCATG
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NcoI

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851 TTCACGCAAA TGGGCGGTAG CCGTGACGG TGGGAGGTCT ATATAAGCAG
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SacII

951 TTGACCTCCA TAGAAGACAC CGGGACCGAT CCAGCCTCCG CGGCCGGGAA
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SphI

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1251 CAACATCTTC TATTGGCTAT ATGCCAATAC TCTGTCTTTC AGAGACTGAC
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1451 TCTCCGGTAG CGGCGGAGCT TCCACATCCG AGCCCTGGTC CCATGCCTCC
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1601 TGGCGGTAGG GTATGTGTCT GAAAATGACC GTGGAGATTG GGCTCGCACG
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NcoI

1801 CGCCCCACCA GACATAATAG CTGACAGACT AACAGACTGT TCCTTTCCAT
GCCCGGTGGT CTGTATTATC GACTGTCTGA TTGTCTGACA AGGAAAGCTA

Sall

NcoI

PmlI BclI EcoRV

1851 CGGTCTTTTC TGCACTCACC GTGCTCGACA CGTGTGATCA GATATCGCGG
CCCAGAAAG ACGTCAGTGG CAGCAGCTGT GCACACTAGT CTATAGCGCC

SphI

EcoRV

1901 CCGCTCTAGC TAGATGCATG CTCGAGCGGC CGCCAGTGTG ATGGATATCT
GGCCAGATCG ATCTACGTAC GAGGTGCGCG GGGTCCACAC TACCTATAGA

NcoI

1951 GCAGAACTCT ATCTTCAGGA TCTCGCCATG GAGGGTCTTA GCCTACTCCA
CGTCTTAAGA TAGAAGTCTT AGAGCGGTAC CTCCAGART CCGATGAGGT

2001 ATTGCCCAGA GATAAATTTT GAAAAAGCTC TTTCTTTCTT TGGGTATCA
TAACGGGTCT CTATTTAAAG CTTTTCGAG AAAGAAACA ACCCAGTAGT

2051 TCTAATTTCA AATGCGCTTT TCCATGCCTT TGGGTGTGTI GACCAACAGC
AGATTAAGT TTYCCGGAAA AGGTACGGAA ACCCAACA CACTCTGCTG

2101 ACTTTAGAAG TAACAGAGAT TGACCAGCTA GTCTGCAAGG ATCATCTTGC
TGAAATCTTC ATTGTCTCTA ACTGGTCTAT CAGACGTTCC TAGTAGAAGC

2151 ATCAACTGAC CAGCTGAAAT CAGTTGGTCT CAACCTCGAG GGGAGCGGAG
TAGTTGACTG GTCGACTTTA CTCACCCAGA GTTGGAGCTC CCCTCGCCTC

EcoRV

2201 TATCTACTGA TATCCCATCT GCGACAAAGC GTTGGGGCTT CAGATCTGGT
ATAGATGACT ATAGGGTAGA CGCTGTTTCG CAACCCCGAA GTCTAGACCA

2251 GTCCCTCCCC AAGTGGTCAG CTATCAAGCA GGAGAATGGG CTGAAATTG
CACGGAGGGG TTCACCAGTC GATACTTCTT CCTCTTACCC GACTTTTAAC

2301 CTACAATCTT GAAATAAAGA AACCAGGACG GACCGAATGC TTACCCCCAC
GATGCTAGAA CTTTATTTCT TTGGCCTGCC CTCGCTTACG AATGCGGGTG

2351 CGCCGGATCG TGTCAAGGC TTTCCAAGGT GCGGCTATCT TCACAAAGCC
GCGGCTACC ACAGTCTCCG AAAGGTTCCA CGGCGATACA AGTCTTTCGG

2401 CAAGGAACCG GGCCTGCCC CGGTGACTAT GCCTTTCACA AGGATCGAGC
GTTCTTGGC CCGGGACGGG CCCACTGATA CGGAAAGTGT TCCTACCTCG

2451 TTCTTCTCTC TATGACAGGC TGGCTTCAAC TGTAATTTAC AGAGGAGTCA
AAAGAAGGAG ATACTGTCCG ACCGAAGTTG ACATTAAATC TCTCTCTAGT

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2501  ATTTTCTGA  CCGGGTAATC  GCATTCCTGA  TATTGGCTAA  ACCAAAGGAA
      TAAAACGACT  CCCCCATTAG  CGTAAGAACT  ATAACCGATT  TGGTTTCCTT
.....
2551  ACGTTCTCTC  AATCACCCCC  CATTGAGAGAG  GCAGCAAACT  ACACCGAAAA
      TCGAAGGAAG  TTAGTGGGGG  GTAAGCTCTC  CGTCGTTTGA  TGTGACTTTT
.....
2601  TACATCAAGT  TACTATGCCA  CATCCTACTT  GGACTACGAA  ATCGAAAAAT
      ATGTAGTTCA  ATGATACGGT  GTAGGATGAA  CCTCATGCTT  TAGCTTTTAA
.....
2651  TTGGTGCTCA  ACACTCCACG  ACCCTTTTCA  AAATTAACAA  TAATACTTTT
      AACCAAGAGT  TGTGAGGTCC  TGGGAAAAGT  TTTAATTGTT  ATATGAAAA
.....
2701  GTTCTCTGG  ACAGGCCCCA  CACGCCTCAG  TTCTTTTCC  AGCTGAATGA
      CAAGAGAGAC  TGTCCGGGGT  GTGCGGAGTC  AAGGAAAAGG  TCGACTTACT
.....
2751  TACCATTCAA  CTTACCAAC  AGTTGAGCAA  CACAACGGG  AAATAATTT
      ATCGTAAGTT  GAAGTCGTTG  TCAACTCGTT  GTGTTGACCC  TTTGATTAAA
.....
2801  GGCACCTAGA  TCGTAATATC  AATGCTGATA  TTGGTGAAAG  GGCTTTTGG
      CCTGTGATCT  ACGATTATAG  TTACGACTAT  AACCACTTAC  CCGAAAAACC
.....
2851  GAAAAATAAA  AAATCTCTCC  GAACAACCTAC  GTGGAGAAGA  GCTGTCTTTC
      CTTTATTTT  TTAGAGAGG  CTTGTTGATG  CACCTCTTCT  CGACAGAAAG
.....
2901  GAACTTTTAT  CGCTCAACGA  GACAGAAGAC  CATGATCCGA  CATCGTCGAG
      CTTTCAAATA  CCGAGTTGCT  CTGCTTCTG  CTACTACGCT  GTAGCAGCTC
.....
2951  AACTACAAG  GGAAGAATCT  CCGACCGGGC  CACCAGGAAG  TATTGGGACC
      TTGATGTTT  CCTTCTTAGA  GGCTGGCCCG  GTGGTCCTTC  ATAAGCCTGG
.....
3001  TGGTTCCAAA  GGATTCCTCT  GGGATGGTTT  CATTGCACGT  ACCAGAAGGG
      ACCAAGGTTT  CCTAAGGGCA  CCCTACCAAA  GTACCGTGCA  TGGTCTTCCC
.....
3051  GAAACAACAT  TGCCCTCTCA  GAATTCGACA  GAAGGTGCAA  GAGTAGATGT
      CTTTGTGTGA  ACGGCAGAGT  CTTAAGCTGT  CTTCAGCTT  CTCATCTACA
.....
3101  GAATACTCAG  GAAACTATCA  CAGAGACAAC  TGCAACAATC  ATAGGCACTA
      CTTATGAGTC  CTTTGATACT  GTCTCTGTTG  ACGTTGTTAG  TATCCGTGAT
.....
3151  ACGGTAACAA  CATGCAGATC  TCCACCATCG  GGACAGGACT  GAGCTCCACC
      TGCCATTGTT  GTACGTCTAG  AGGTGCTAGC  CCTGTCTTGA  CTCGAGGTCT
.....

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NcoI

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3201  CAATCTCTGA  GTTCTCACC  GACCATGCCA  CCAAGCCCTG  AACTCAGAC
      GTTAGGACT  CAAGGAGTGG  CTGGTAACGT  GGTTCGGGAC  TCTGACTCTG
.....
3251  CTCCACAACC  TACACACCAA  AACTACCAGT  GATGACCACC  GAGGAACCAA
      GAGGTGTTGG  ATGTGTGCTT  TTGATGGTCA  CTACTGCTGG  CTCCTTGGTT
.....
3301  CACCAACCAC  GAGAACTCT  CCTGGCTCAA  CAACAGAAGC  ACCCACTCTC
      GTGTGCTGG  CTCTTTGAGA  GGACCGAGTT  GTGTCTTTC  TGGGTGAGAG
.....
3351  ACCACCCACG  AGAATATAAC  AACACCGGTT  AAACTGTTT  GGGCACAAGA
      TGGTGGGGTC  TCTTATATTG  TTGTCGCCAA  TTTTGACAAA  CCCGTGTTCT
.....

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3401 GTCCACAAGC AACGGTCTAA TAACCTCAAC AGTAACAGGT ATTCTTGGGA
CAGGTGTTCC TTGCCAGATT ATTGAAGTTG TCATTGTCCA TAAGAACCCT

3451 GCCTTGGACT TCGAAAACGC AGCAGAAGAC AAGTTAACAC CAGGCCACCG
CGGAACCTGA AGCTTTTGCG TCGTCTTCTG TTCAATTGTG GTCCTGGTGC

3501 GGTAATGCA ATCCCAACTT ACACTACTGG ACTGCCAAG AACAAACATAA
CCATTTACGT TAGGGTTGAA TGTGATGACC TGACGTGTC TTGTGTATT

BamHI

3551 TGCTGCTGGG ATTGCCTGGA TCCCGTACTT TCCACCGGGT GCAGAAGGCA
ACGACGACCC TAACGGACCT AGGGCATGAA ACCTGGCCCA CGTCTTCCGT

3601 TATCACTGA AGGCCTTATG CACAACCAA ATGCCTTAGT CTGTGGACTC
ATATGCTACT TCCGGAATAC GTGTTGGTTT TACGGAATCA CACACCTGAG

3651 AGACAACCTG CAAATGAAAC AACTCAAGCT CTCCAGCTTT TCTTAAGGGC
TCTGTTGAAC GTTACTTTG TGAAGTTCCA GACGTCGAAA AGAATCCCG

3701 CACGACGGAG CTGCGGACAT ATACCATACT CAATAGGAAG GCCATAGATT
GTGCTGCCTC GACGCCTGTA TATGGTATGA GTTATGCTC CGGTATCTAA

BamHI

3751 TCCTTCTGCG ACGATGGGCC GGGACATGTA GGATCCTGGG ACCAGATGT
AGGAAGACCC TGCTACCCCG CCTGTACAT CCTAGGACCC TGGTCTAACA

3801 TGCATTGACC CACATGATTG GACCAAAAC ATCACTGATA AAATCAACCA
ACGTAACCTG GTGTACTAAC CTGGTTTTG TAGTGACTAT TTTAGTTGGT

3851 AATCATCCAT GATTTTCATC ACAACCTTT ACCCAATCAG GATAATGATG
TTAGTAGGTA CTAAGTAGC TGTGGGAAA TGGGTTAGTC CTATTACTAC

BamHI

3901 ATAATTGGTG GACGGGCTGG AGACASTGGA TCCCTGCAGG AATAGGCATT
TATTAACCAC CTGCCCGACC TCTGTCACCT AGGGACGTCC TTATCCGTAA

3951 ACTGGAATTA TTATTGCAAT CATGCTCTT CTTTGGCTCT GCAAGCTGCT
TGACCTTAAT AATAACGTTA GTAACGAGAA GAAACCCAGA CGTTCGACGA

BamHI

4001 TTGTTGAATA TCAGAATTCC AGCACTGGCG GCCGTACTA GTGGATCCGA
AACAACTTAT AGTCTTAAGG TCGTGACCGC CGGCAATGAT CACCTAGGCT

NarI

BamHI

XbaI

KasI

BamHI

4051 GCTCGGATCC AAGCTCTAGA CCAGGCGCCT GGATCCAGAT CTGCTGTGCC
CGAGCCTAGG TTCCAGATCT GGTCCGCGGA CCTAGGTCTA GACGACACCG

4101 TTCTAGTGG CAGCCATCTG TTGTTTGCCC CTCCCCGTC CCTTCCTGA
AAGATCAACC GTCGGTAGAC AACAAACGG GAGGGGGCAC GGAAGGAAC

4151 CCCTGGAAGG TGGCACTCCC ACTGTCCCTT CCTAATAAAA TGAGGAAATT
GGCACCTTCC ACGGTGAGGG TGACAGGAAA GGATTATTTT ACTCCTTTAA

4201 GCATCGCATT GTCTGAGTAG GTGTCAATCT ATTCTGGGGG GTGGGGTGGG
CGTAGCGTAA CAGACTCATC CACAGTAAGA TAAGACCCCC CACCCACCC

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SphI
.....

4251 CCAGCACAGC AAGCGGAGG ATTGGGAAGA CAATACCAGG CATGCTGGGG
CGTCGTGTCG TTCCTCTCC TAACCTTCT GTTATCGTCC GTACGACCC

.....
KpnI
.....

4301 ATGCGGTGGG CTCTATGGGT ACCCAGGTGC TGAAGAATTG ACCCGGTTC
TACGCCACCC GAGATACCCA TGGGTCCACC ACTTCTTAAC TGGGCCAAGG

4351 TCCTGGGCCA GAAAGAAGCA GGCACATCCC CTTCTCTGTG ACACACCTG
AGGACCCGGT CTTTCTTCGT CCGTGTAGCG GAAGAGACAC TGTGTGGGAC

4401 TCCACGCCCC TGGTCTTAG TTCCAGCCCC ACTCATAGGA CACTCATAGC
AGGTGCGGGG ACCAAGAATC AAGGTGCGGG TGAGTATCCT GTGAGTATCG

4451 TCAGGAGGCG TCCGCTTCA ATCCCACCCG CTAAAGTACT TGGAGCGGTC
AGTCTCCCG AGCGGAAGT TAGGGTGGCG GATTTCATGA ACCTCGCCAG

4501 TCTCCCTCCC TCATCAGCCC ACCAAACCAA ACCTAGCCTC CAAGAGTGGG
AGAGGGAGGG AGTAGTCGGG TGGTTGGTT TGGATCGGAG GTTCTCACC

4551 AAGAAATTAA AGCAAGATAG GCTATTAAAT GCAGAGGGAG AGAAATGCC
TCTTTAATT TCGTTCTATC CGATAATTCA CGTCTCCCTC TCTTTACGG

.....
XbaI
.....

4601 TCCACATGT GAGGAAGTAA TCAGAGAAAT CATAGATTT CTTCGGCTC
AGGTTGTACA CTCCTTCATT ACTCTCTTA GTATCTTAA GAAGCGAAG

4651 CTCGCTCACT GACTCGCTGC GCTCGGTCTG TCGGCTCGG CGAGCGGTAT
GAGCGAGTGA CTGAGCGAGC CGAGCCAGCA AGCCGACGCC CTCGCCATA

4701 CAGCTCACTC AAAGGCGGTA ATACGGTTAT CCACAGAATC AGGGGATAAC
GTCGAGTGAG TTTCCGCCAT TATGCCAATA GGTGTCTTAG TCCCTATTG

4751 GCAGGAAGA ACATGTGAGC AAAAGGCCAG CAAAAGGCCA GCAACCGTAA
CGTCTTTCT TGTACACTCG TTTCCGGTC GTTTCCGGT CCTTGGCATT

4801 AAAGCCCGCG TTGCTGGCGT TTTCCATAG GCTCCGCCCC CCTGACGAGC
TTTCCGGCCG AACGACCGCA AAAAGGTATC CGAGGCGGGG GGACTGCTCG

4851 ATCACAATAA TCGACGCTCA AGTCAGAGCT GCGGAAACCC GACAGGACTA
TAGTGTTTT AGCTCCGAGT TCAGTCTCCA CCGCTTTGG CTGTCTGAT

4901 TAAAGATACC AGGCGTTTCC CCTGGAAGC TCCCTCGTGC GCTCTCCTGT
ATTCTATGC TCCGCAAGG GGGACCTTCG AGGAGCACG CGAGAGGACA

4951 TCCGACCTCG CCGCTTACCG GATACCTGTC CGCCTTTCTC CCTTCGGGA
AGGCTGGGAC GGCGAATGCC CTATGGACAG GCGGAAAGAG GGAAGCCCTT

5001 GCGTGGCGCT TTCTCAATCC TCACGCTGTA GGTATCTCAG TTCGGTGTAG
CGCACCGCGA AAGAGTTACG AGTCCGACAT CCATAGAGTC AAGCCACATC


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5051  GTCGTTCCCT  CCAAGCTGGG  CTGTGTGCAC  GAACCCCCCG  TTCAGCCCGA
      CAGCAAGCGA  GGTTCGRCCC  GACACACGTG  CTTGGGGGGC  AAGTCGGGCT
.....
5101  CCGCTGCCCC  TTATCCGGTA  ACTATCGTCT  TGAATCCAAC  CCGGTAAGAC
      GGCGACGGGG  AATAGGCCAT  TGATAGCAGA  ACTCAGGTTG  GGCCATTCTG
.....
5151  ACGACTTATC  GCCACTGGCA  GCAGCCACTG  GTAACAGGAT  TAGCAGACGG
      TGCTGAATAG  CGGTGACCGT  CGTCGGTGAC  CATTGTCTTA  ATCGTCTCGC
.....
5201  AGGTATGTAG  GCGGTGCTAC  AGAGTTCTTG  AAGTCGTGGC  CTAACTACGG
      TCCATACATC  CGCCACGATG  TCTCAAGAAC  TTCACCACCG  GATTGATGCC
.....
5251  CTACACAGAG  AGGACAGTAT  TTGGTATCTG  CGCTCTGCTG  AAGCCAGTTA
      GATGTGATCT  TCCTGTGATA  AACCATAGAC  GCGAGACGAC  TTCGGTCAAT
.....
5301  CCTTCGGAAA  AAGAGTTGGT  AGCTCTTGAT  CCGGCAACA  AACCACCGCT
      GCAAGCCTT  TTCTCAACCA  TCGACAACCT  GCGCGTTTGT  TTGGTGGCGA
.....
5351  GGTAGCGGTG  GTTTTTTTGT  TTGCAAGCAG  CAGATTACGC  GCAGAAAAAA
      CCAATGCCAC  CAAAAAACA  AACGTTCTGC  GTCTAATGCC  CGTCTTTTTT
.....
5401  ACGATCTCAA  GAAGATCCTT  TGATCTTTTC  TACGGGGTCT  CACGCTCAGT
      TCCTAGAGTT  CTTCTAGGAA  ACTAGAAAAG  ATGCCCCAGA  CTGCGAGTCA
.....
5451  GGAACGAAAA  CTCACGTTAA  GGGATTTTGG  TCATGAGATT  ATCAAAAAGG
      CCTTGCTTTT  GAGTCCAATT  CCCTAAAACC  AGTACTCTAA  TAGTTTTTCC
.....
5501  ATCTTCACCT  AGATCCTTTT  AAATTAAAAA  TGAAGTTTAA  AATCAATCTA
      TAGAAGTGG  TCTAGGAAA  TTTAATTTTT  ACTTCAAAAT  TTAGTCAAGT
.....
5551  AAGTATATAT  GAGTAAACTT  GGTCTGACAG  TTACCAATGC  TTAATCAGTG
      TTCATATATA  CTCATTTGAA  CCAGACTGTC  AATGGTTACG  AATTAGTCAC
.....
5601  AGGCACCTAT  CTCAGCGATC  TGTCTATTTT  GTTCATCCAT  AGTTGCCTGA
      TCCGTGGATA  CAGTCGCTAG  ACAGATAAAG  CAAGTAGGTA  TCAACGGACT
.....
5651  CTCCGGGGGG  GGGGGGGCGT  GAGGTCTGCC  TCGTCAAGAA  GGTGTTGCTG
      GAGGCCCCC  CCCCCCGCGA  CTCCAGACGG  AGCACTTCTT  CCACAACGAC
.....
5701  ACTCATACCA  GGCCTGAATC  GCCCCATCAT  CCAGCCAGAA  AGTGAGGCAG
      TGAGTATGGT  CCGGACTTAG  CGGGGTAGTA  GGTGCGTCTT  TCACTCCCTC
.....
5751  CCACGGTTGA  TGAGAGCTTT  GTTGTAGCTG  GACCAATTGG  TGATTTTGAA
      GGTGCCAACT  ACTCTCGAAA  CAACATCCAC  CTGGTCAACC  ACTAAAACCT
.....
5801  CTTATGCTTT  GCCACGGAAC  GGTCTCGGTT  GTGCGGAAGA  TCCGTGATCT
      GAAAACGAAA  CGGTGCCTTG  CCAGACGCAA  CAGCCCTTCT  ACGCACTAGA
.....
5851  GATCCTTCAA  CTCAGCAAAA  GTTCGATTTA  TTCAACAAAG  CCGCCGTCCT
      CTAGGAAGTT  GAGTCGTTTT  CAAGCTAAAT  AAGTTGTTTC  GCGGCGAGGG
.....
5901  GTCAAGTCAG  CGTAATGCTC  TGCCAGTGTT  ACAACCAATT  AACCAATTCT
      CAGTTCAGTC  GCATTACGAG  ACGGTCACAA  TGTGCTTAA  TTGGTTAAGA
.....
5951  GATTAGAAAA  ACTCATCGAG  CATCAATGCA  AACTGCAATT  TATTCATATC
      CTAATCTTTT  TGAGTAGCTC  GTAGTTTACT  TTGACGTTAA  ATAAGTATAG
.....

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6001 AGGATTATCA ATACCATATT TTGAAAAAG CCGTTTCGT AATGAAGGAG
TCCTAATAAT TATGGTATAA AACTTTTTTC GCCAAGACA TTACTTCCTC

6051 AAAACTCACC GAGGCAGTTC CATAGCATGG CAAGATCCTG GTATCGGTCT
TTTTGAGTGG CTCGCTCAAG GTATCCTACC GTTCTAGGAC CATAGCCAGA

6101 GCGATTCEGA CTCGTCCAAC ATCAATACAA CCTATTAAAT TCCCCTCGTC
CGCTAAGGCT GAGCAGGTGG TAGTTATGTT CGATAATTAA AGGGGAGCAG

6151 AAAAATAAGG TTATCAAGTG AGAAATCACC ATGAGTGACG ACTGAATCCG
TTTTTATTCC AATAGTTCAC TCTTAGTGG TACTCACTGC TGACTTAGGC

HindIII

6201 GTGAGAATGG CAAAAGCTTA TGCATTTCCT TCCAGACTTG TCAACAGGC
CACTCTTACC GTTTTCGAAT ACGTAAAGAA AGGTCTGAAC AAGTTGTCCG

6251 CAGCCATTAC GCTCGTCAAC AAAATCACTC GCATCAACCA AACCGTTACT
GTCCGTAATG CGAGCAGTAG TTTTAGTGAG CCTAGTTGGT TTGGCAATAA

PvuI

6301 CATTCTGAT TCCGCTGAG CGAGACGAA TACCGGATCG CTGTTAAAG
GTAAGCACTA ACGCGGACTC GCTCTGCTTT ATGCGCTAGC GACAATTTTC

6351 GACAATTACA AACAGGAATC GAATGCAACC GCGCCAGGAA CACTGCCAGC
CTGTTAATGT TTGCTCTAG CTTACGTTGG CCGCTCCTT GTGACGGTGG

6401 GCATCAACAA TATTTTCACC TGAATCAGGA TATTCTTCTA ATACCTGGAA
CGTAGTTGTT ATAAAGTGG ACTTAGTCCT ATAAGAAGAT TATGGACCTC

6451 TGCTGTTTC CCGGGGATCG CAGTGGTGG TACCATGCA TCATCAGGAG
ACGACAAAAG GCGCCCTAGC GTCACCACTC ATTGGTACGT AGTAGTCCCT

6501 TACGGATAAA ATGCTTGATG GTCGGAAGAG GCATAAATC CGTCAGCCAG
ATCCCTATTT TACGAACCTAC CAGCCTTCTC CGTATTTAAG GCAGTCGGTC

6551 TTACTCTGA CCATCTCATC TGTAACATCA TTGGCAACGC TACCTTTGCC
AAATCAGACT GGTAGAGTAG ACATTGTAGT AACCGTTGGG ATGCAACGG

ClaI

6601 ATGTTTCAGA AACAACTCTG GCGCATCGGG CTCCCATAC AATCGATAGA
TACAAAGTCT TTGTTGAGAC CCGTAGCCC GAAGGGTATG TTACCTATCT

6651 TTGTCGCCACC TGATTGCCCG ACATTATCGC GAGCCCATTT ATACCATAT
AACAGCGTGG ACTAACGGGC TGTAATAGCG CTCGGGTAAA TATGGGTATA

6701 AAATCAGCAT CCATGTTGGA ATTTAATCGC GGCCTCGAGC AACAGTTTC
TTTAGTCGTA GGTACAACCT TAAATTAGCG CCGGAGCTCG TTCTGCAAG

6751 CCGTTGAATA TGGCTCATAA CACCCCTTGT ATTACTGTTT ATGTAAGCAG
GGCACTTAT ACCGAGTATT GTGGGGAACA TAATGACAAA TACATTGCTC

6801 ACAGTTTAT TGTTCAATGAT CATATATTTT TATCTTGTC AATGTAACAT
TGCAAAATA ACAAGTACTA CTATATAAAA ATAGAACACG TTACATTGTA

DraIII

6851 CAGAGATTTT GAGACACAAC GTGGCTTTCC CCCCCCCCCC ATTATTGAAG
 CTCTCTAATA CTCTGTGTTG CACCCAAAGG GCGGGGGGGG TAATAACTTC

6901 CATTATACAG GGTATTGTC TCATGAGCGG ATACATATTT GAATGTATT
 GTAAATAGTC CCAATAACAG AGTACTCGCC TATGTATATA CTACATAAA

6951 AGAAAAATAA ACAAAATAGGG GTTCCGCCCA CATTCCCCG AAAAGTGCCA
 TCTTTTATT TGTATTCCG CAAGGCCCGT GTAAAGGGGC TTTTCACGGT

7001 CCTGACGTCT AAGAAACCAT TATTATCATG ACATTAACCT ATAAAAATAG
 CCACTGCAGA TTCTTTGGTA ATAATAGTAC TGTAAATGGA TATTTTATC

7051 CCGTATCAG AGCCCTTTT CTC
 CGCATAGTGC TCCGGGAAG CAG

SEQUENCE LISTING ID NO: 3

pVR 1012-GP(Z)

General Description

DNA pVR 1012-GP(Z)
 Local object
 Created: 09/15/98 05:06PM
 Last Modification Date: ? (no data)
 length: 7285 bp
 storage type: Basic
 form: Circular

Comments

Restriction Map

DraIII: 1 site CACNNNGTG
 GTGNNAC
 HindIII: 1 site AAGCTT
 TTCGA
 HpaI: 1 site GTTAC
 CAATTC
 KsaI: 1 site GCGGCC
 GCGCGG
 NarI: 1 site GCGGCC
 CCGCGG
 NotI: 1 site CCGGCCCG
 CGCGCGG
 PmlI: 1 site CACGTG
 GTGCAC
 PvuI: 1 site CGATCG
 GGTAGC
 SacII: 1 site CCGCGG
 GCGGCC
 XbaI: 1 site TCTAGA
 AGATCT
 XhoI: 1 site CTCGAG
 GAGCTC
 EcoRV: 2 sites GATATC
 CTATAG
 NcoI: 2 sites CCATCG
 GGTACG
 NdeI: 2 sites CATATG
 GTXTAC
 SphI: 2 sites CCATCG
 CGTACG

Functional Map

CDS (4 signals)

CMV IE 5' UT

Start: 886 End: 1129

CMV IE INT

Start: 1130 End: 1840

TbGH

Start: 4302 End: 4854

Kan^r

Start: 6350 End: 6972 (Complementary)

Misc_featur (2 signals)

CMV enhancer

Start: 248 End: 885

GP(Z)

Start: 1870 End: 4301

Annotations

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1  TCGCCCGTTT CGGTGATGAC GGTGAAAACC TCTGACACAT GCAGCTCCCG
   AGCGCGCAAA GCCACTACTG CCACTTTTGG AGACTGTGTA CGTCGAGGGC
.....
51  GAGACGGTCA CAGCTTGTCT GTAAACGGAT GCCGGGAGCA GACAAGCCCCG
   CTCGCCCAGT GTCGACAGA CATTCGCCTA CGGCCCTCGT CTGTTGGGGC
.....
101 TCAGGGCGCG TCAGCGGGTG TTGGCGGGTG TCGGGGCTGG CTTAACTATG
   AGTCCCGCGC AGTCGCCCCAC AACCGCCAC AGCCCCGACC GAATTGATAC
.....

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NdeI

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151 CCGCATCAGA GCAGATTGTA CTGAGAGTGC ACCATATGCG GTGTGAAATA
   CCGTAGTCT CGTCTAACAT GACTCTCAGG TGGTATACGC CACACTTTAT
.....
201 CCGCACAGAT GCGTAAGGAG AAAATACCGC ATCAGATTGG CTATTGGCCA
   GCGGTGCTTA CGCATTCTTC TTTTATGGCG TAGTCTAACG GATAACCGGT
.....
251 TTGCATACGT TGTATCCATA TCATAATATG TACATTTATA TTGGCTCATG
   AACGTATCCA ACATAGGTAT AGTATTATAC ATGTAAATAT AACCGAGTAC
.....
301 TCCAACATTA CCGCCATGTT GACATTGATT ATTGACTAGT TATTAATAGT
   AGGTTGTAAT GCGGTACAA CTGTAACATA TAACTGATCA ATAAATATCA
.....
351 AATCAATTAC GGGGTCATTA GTTCATAGCC CATATATGGA GTTCCGCGTT
   TTAGTTAATG CCCCAGTAAT CAAGTATCGG GTATATACCT CAAGCGCGAA
.....
401 ACATAACTTA CCGTAATTCG CCCGCCCTGG TCACCGCCCA ACGACCCCGG
   TGTATTGAAT CCCATTACG GCGCGGACCG ACTGGCGGGT TGCTGGGGGC
.....
451 CCCATTGACG TCAATAATGA CGTATGTTCC CATAGTAACG CCAATAGGGA
   GGGTAATGCG AGTTATTACT GCATACAAGG GTATCATGCG GGTATCCCT
.....
501 CTTTCCATTG ACGTCAATGG GTGGAGTATT TACGGTAAAC TGCCCACTTG
   GAAAGGTAAC TGCAGTTACC CACCTCATAA ATGCCATTG ACGGGTGAAC
.....

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NdeI

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551 GCAATACATC AAGTGTATCA TATGCCAAGT ACGCCCCCTA TTGACGTCAA
   CGTCATGTAG TTCACATAGT ATACGGTTCA TCGGGGGGAT AACTGCAGTT
.....
601 TGACGGTAAA TGGCCCCCCT GGCATTATGC CCAGTACATG ACCTTATGGG
   ACTGCCATT ACCCGCGCGA CCGTAATACG GGTCAATGAC TGAATACCC
.....

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NcoI

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651 ACITTCCTAC TTGGCAGTAC ATCTACGTAT TAGTCATCGC TATTACCATG
   TGAAGGATG AACCGTCATG TAGATGCATA ATCAGTAGCG ATAATGGTAC
.....

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NcoI

```

701 GTGATCGCGT TTTGGCAGTA CATCAATGGG CGTGGATAGC GGTTTGACTC
   CACTACGCCA AAACCGTCAT GTAGTTACCC GCACCTATCG CCAAACTGAG
.....
751 ACCGGGATTT CCAAGTCTCC ACCCCATTGA CGTCAATGGG AGTTTGTTTT
   TGGCCCTAAA GGTTCAGAGG TGGGGTAACG GCAGTTACCC TCAAACAAA
.....
801 GGCACCAAAA TCAACGGGAC TTTCCAAAAT GTCGTAACAA CTCCGCCCCA
   CCGTGGTTTT ACTTGGCCCTG AAAGGTTTTA CAGCATTGTT GAGGCGGGGT
.....

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851 TTGACGCAA TGGCGGOTAG GCGGTACGG TGGGAGGTCT ATATAAGCAG
AACTGCGTTT ACCCGCCATC CGCACATGCC ACCCTCCAGA TATATTGCTC

901 AGCTCGTTTA GTGAACCGTC AGATCGCCTG GAGACGCCAT CCACCGTGT
TCGAGCAAAT CACTTGGCAG TCTAGCCGAC CTCTCGGTA GTTGGACAA

SacII

951 TTGACCTCCA TAGAAGACAC CGGACCGAT CCAGCCTCCG CGGCGGGAA
AACTGGAGGT ATCTTCTGTG GCCCTGGCTA GGTGGGAGGC GCCGCGCCTT

1001 CCGTGCATTG GAACCGGAT TCCCCGTGCC AAGAGTGACG TAAGTACCGC
GCCACGTAACT CTTCGCGCTA AGGGGCACGG TTCTCACTGC ATTCAATGGCG

SphI

1051 CTATAGACTC TATAGGCACA CCCCTTTGGC TCTTATGCAT GCTATACTGT
GACATCTGAG ATATCCGTGT GCGGAAACCG ACAATACGTA CGATATGACA

1101 TTTTGGCTTG CGGCCTATAC ACCCCCGCTT CTTATGCTA TAGGTGATGG
AAAACCGAAC CCCGGATATG TGGCGGCGAA GGAATACGAT ATCCACTACC

1151 TATAGCTTAG CCTATAGGTG TGGGTATTG ACCATTATG ACCACTCCCC
ATATCGAATC GGATATCCAC ACCCAATAAC TGGTAATAAC TGGTGAGGGG

1201 TATTGGTGAC GATACTTTC AATACTAATC CATACATGG CTCTTTGCCA
ATAACCACTG CTATGAAAGC TAATGATTAG GTATTGTACC GAGAAACGGT

1251 CAACTATCTC TATTGGCTAT ATGCCAATAC TCTGTCTTC ACAGACTGAC
GTTGATAGAG ATAACCGATA TACGGTTATG AGACAGGAAG TCTCTGACTC

1301 ACCGACTCTC TATTTTACA GGATGGGGTC CCATTATTG TTTACAAAT
TGCTTGAGAC ATAAAAATGT CCTACCCAG GGTAAATAAT AAATGTTAA

1351 CACATATACA ACAACGCCGT CCCCCGTGCC CGCAGTTTTT ATTAACATA
GTGTATATGT TGTTCGGCA GGGGGCACGG CGGTCAAAA TAATTTGTAT

1401 CCGTGGGATC TCCACCGGAA TCTCGGGTAC GTGTTCGGGA CATGGGCTCT
CGCACCCCTAG AGGTGCGCTT AGAGCCCATG CACAAGGCCT GTACCCGAGA

1451 TCTCCGCTAG CGCGGGAGCT TCCACATCCG AGCCCTGGTC CCATGCCTCC
AGAGGCCATC CGCGCCTCGA AGGTGTAGGC TCGGGACCAG GGTACGGAGG

1501 AGCGGCTCAT GCTCGCTCGG CAGCTCCTTG CTCCTAACAG TGGAGCCAG
TCCCGGAGTA CCAGCGAGCC GTCGAGGAAC GAGGATTGTC AACTCGGTC

1551 ACTTAGGCAC AGCACAATGC CCACCACCAC CAGTGTGCGG CACAAGGCG
TGAATCCGTG TCGTGTACG GGTGGTGGTG GTCACACGGC GTGTTCCGGC

1601 TGGCGGTAGG GTATGTGTCT GAAAATGAGC GTGGAGATTG GGCTCGCAG
ACCCCATCC CATAACAGA CTTTACTCG CACCTCTAAC CCGAGCGTC

1651 GCTACCGCAG ATGGAAGACT TAAGCCAGCG CCAGAAGAAG ATGCAGGCAG
CGACTGCGTC TACCTTCTGA ATTCCGTCCG CGTCTTCTTC TACGTCCGTC

1701 CTGAGTCTT GTATTCTGAT AAGAGTCAGA GGTAACTCCC GTTCCGGTGC
GACTCAACA CATAAGACTA TTCTACGCT CCATTGAGGG CAACGCCAG

RpaI

1751 TGTTAACGGT GGAGGCGAGT GTAGTCTGAG CAGTACTCGT TGCTGCCGCG
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NcoI

1801 CGCGCCACCA GACATAATAG CTGACAGACT AACAGACTGT TCCTTTCCAT
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NcoIPmlIEcoRVNotI

1851 GGGTCTTTTC TGCAGTCACC GTCCTCGACA CCGTGTGATCA GATATCGCGG
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NarINotI XbaIKasI

1901 CCGCTCTAGA CCAGGCGCCT GGATCGATCC GCGATGAAGA TTAAGCCGAC
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1951 AGTCAGCGTA ATCTTCATCT CTCCTAGATT ATTTGTTTC GACAGTAGGG
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2001 GTCGTCAGGT CCTTTTCAAT CGTGTAAACA AATAAACTC CACTAGAAGG
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2051 ATATTGIGGG GCAACAACAC AATGGGCGTT ACAGGAATAT TCCAGTTACC
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2101 TCGTGATCGA TTCAAGAGGA CATCATCTTT TCTTTGGGTA ATTATCCTTT
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2151 TCCAAAGAAC ATTTTCCATC CCACTTCGAG TCATCCACAA TAGCACATTA
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2201 CAGGTTAGTC ATGTCGACAA ACTAQTITGT CCGTACAAAC TGTCATCCAC
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2251 AATCAATTG AGATCAGTTG GACTGAATCT CGAAGCGAAT GGAGTGGCAA
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2301 CTGACGTGCC ATCTGCAACT AAAAGATGGG CCTTCAGGTC CGGTGTCCCA
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2351 CCAAGGTGG TCAATTATGA ACCTGGTGAA TGGGCTGAAA ACTGCTACAA
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2401 TCTTGAARTC AAAAAACCTG ACGGCAATGA GTGTCTACCA GCAGCGCCAG
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2451 ACGGGATTTC GGGCTTCCCC CGGTGCCGCT ATGTCCACAA AGTATCAGGA
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2501 ACGGACCCCT GTCCCGGAGA CTTTGCTTC CATAAAGAGG GTGCTTCTT
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2551 CCGTATGAT CGACTTCCTT CCACAGTTAT CTACCGAGGA ACGACTTTTCG
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2601 CTGAAGGTGT CGTTGCATTT CTGATACTGC CCCAAGCTAA GAAGGACTTC
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2651 TTCAGCTCAC ACCCCTTGAG AGAGCCGGTC AATGCAACGG AGGACCCGTC
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EcoRV

2701 TAGTGGCTAC TATTCTACCA CAATTAGATA TCAGGCTACC GGTTTTGGAA
ATCAACGATG ATAAGATGGT GTTAATCTAT AGTCCGATCG CCAAAACCTT

2751 CCAATGAGAC AGAGTACTTG TTCGAGGTTG ACAATTTGAC CTACGTCCAA
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2801 CTGGAATCAA GATTACACAC ACAGTTTCTG CTCGAGCTGA ATGAGACAAT
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3001 TGTATCAAAC GGAGCCAAAA ACATCAGTGG TCAGACTCCG GCGCGAACTT
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3051 CTTCCGACCC ACGGACCAAC ACAACAACCTG AAGACCACAA AATCATGGCT
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3151 TCCAGTGTGG CATCTAACAA CCCTTGCCAC AATCTCCAG AGTCCCAAT
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3201 CCCTCACAAC CAACCCAGGT CCGGACAACA GCACCCATAA TACACCCGTG
GGCAGTGTG GTTGGTCCA GGCCTGTTGT CGTGGGTATT ATCTGGGCAC

3251 TATAAACTTG ACATCTCTGA CCAACTCAA GTTGAACAAC ATCACCOCAG
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3301 AACAGACAC GACAGCACAG CCTCCGACAC TCCCTCTGCC ACGACCGCAG
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3351 CCGGACCCCC AAAAGCAGAG AACACCAACA CGAGCAAGAG CACTGACTTC
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3401 CTGGACCCCG CCACCACAAC AAGTCCCAA AACCACAGCG AGACCGCTGG
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SphI

4501 GGCATGCTGG GGATCGGGTG GCTCTATGG GTACCCAGGT GCTGAACAAT
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4551 TGACCCCGTT CCTCCTGGGC CAGAAAGAAG CAGGCACATC CCCTTCTCTG
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4601 TGACACACCC TGTCCACGCC CCTGGTTCTT AGTCCAGCC CCACTCATAG
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4651 GACACTCATA GCTCAGGAGG GCTCCGCCCT CAATCCCACC CGCTAAAGTA
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4701 GTTGGAGCGG TCTCTCCCTC CCTCATCAGC CCACCAAAACC AAACCTAGCC
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4751 TCCAAGAGTG GGAAGAAATT AAAGCAAGAT AGGCTATTAA GTGCACAGGG
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 5851 ATAGTTGCGT GACTCCGCGG GGGGGGGGCG CTGAGGTCTG CCTCGTGAAG
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 6201 TTTATTCATA TCAGGATTAT CAATACCATA TTTTGAATA AGCCGTTTCT
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6251 GTAATGAAGG AGAAACTCA CCGAGCCAGT TCCATAGGAT GGCAAGATCC
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6301 TGGTACGGT CTGCGATTC GACTCGTCCA ACATCAATAC AACCTATTAA
ACCATAGCCA GACGCTAAGG CTGACCAGGT TGTAGTTATG TTGGATAATT

6351 TTCCCCCTCG TCAAAAATAA GGTATCAAG TCAGAAATCA CCATGAGTGA
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HindIII

6401 CGACTGAATC CCGTGACAAT GGCAAAAGCT TATGCATTTC TTTCCAGACT
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6451 TGTTC AACAG GCCAGCCATT ACGCTCGTCA TCAAAATCAC TCGCATCAAC
ACAAGTTGTC CCGTCCGTA TCGGAGCAGT AGTTTTAGTG ACGGTAGTTG

PvuI

6501 CAAACCGGTA TTCATTCGTG ATTGCGCCTG AGCGAGACGA AATACGCGAT
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PvuI

6551 CGCTGTTAAA AGGACAATTA CAAACAGGAA TCGAATGCAA CCGGCGCAGG
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6601 AACACTGCCA GCCCATCAAC AATATTTTCA CCTGAATCAG CATACTCTTC
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6801 CCTACCTTTG CCATGTTTCA GAAACAACTC TGGCGCATCG GGCTTCCCAT
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6851 ACAATCGATA GATTGTCGCA CCTGATTGCC CGACATTATC CCGAGCCCAT
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XhoI

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XhoI

6951 GCAAGACGTT TCCCGTTCAA TATGGCTCAT AACACCCCTT GTATTACTGT
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7001 TTATGTAACC AGACAGTTT ATTGTTTATC ATGATATATT TTTATCTTGT
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DraIII

7051 GCAATGTAAC ATCAGAGATT TTGAGACACA ACGTGGCTTT CCCCCCCCCC
 CGTTACATTG TAGTCTCTAA AACTCTGTGT TGCACCGAAA CCGGGGGGGG

7101 CCATTATTGA AGCATTATC AGCGTATTG TCTCATGAGC GGATACATAT
 GGTAACTAACT TCGTAAATAG TCCCAATAAC AGAGTACTCG CCTATGTATA

7151 TTGAATGTAT TTAGAAAAAT AAACAAATAG GGGTCCCGC CACATTCCC
 AACTTACATA AATCTTTTFA TTTGTTTATC CCCAAGGCGC GTGTAAAGGG

7201 CGAAAAGTGC CACCTGACGT CTAAGAAACC ATTATTATCA TGACATTAAAC
 GCTTTTCACG GTGGACTGCA GATTCTTTGG TAATAATAGT ACTGTAATTG

7251 CTATAAAAAAT AGGCCTATCA CGAGGCCCTT TCGTC
 GATATTTTFA TCCGCATAGT GCTCCGGGA ACCAG

pvr 1012-SG2(Z)

General Description

DNA pvr 1012-SGP(Z)
 Local object
 Created: 09/14/98 04:29PM
 Last Modified: 09/15/98 04:50PM
 length: 7272 bp
 storage type: Basic
 form: Circular

Comments

Restriction Map

DraIII: 1 site CACNNNGTG
 GTGNNNCAC

 HindIII: 1 site AAGCTT
 TTCGTA

 HpaI: 1 site GTTAAC
 CAATTC

 KpnI: 1 site GGTACC
 CCAATG

 NotI: 1 site GCGGCCGC
 CGCGGGCG

 PmlI: 1 site CACGTG
 GTGCAC

 PvuI: 1 site CGATCG
 GCTAGC

 SacI: 1 site CCGCGG
 GCGGCC

 XbaI: 1 site TCTAGA
 AGATCT

 XhoI: 1 site CTCGAC
 GAGCTC

 EcoRV: 2 sites GATATC
 CTATAG

 NcoI: 2 sites CCATGG
 GGTACC

 NdeI: 2 sites CATATC
 GTATAC

 SphI: 2 sites GCATCG
 CATTAC

Functional Map

CDS (4 signals)

CMV IE 5' UT

Start: 886 End: 1129

CMV IE INT

Start: 1130 End: 1840

TbGH

Start: 4289 End: 4841

Kanr

Start: 6337 End: 6959 (Complementary)

Misc_feature (2 signals)

WO 99/37331

PCT/US99/01382

CMV enhancer

Start: 248 End: 885

SGP(Z)

Start: 1870 End: 4288

Annotations

1 TCCCGCGTTT CGGTGATGAC GGTGAAAACC TCTGACACAT GCAGCTCCCG
 ACCGCGCAAA GCGCTACTG CCACTTTTGG AGACTGTGTA CGTCGAGGGC

 31 GACACGGTCA CAGCTTGCT GTAGCGGAT CCCGGGAGCA GACAAGCCCG
 CTCGCCAGT GTCGAACAGA CATTGCGCTA CGGCCCTCGT CTGTTCCGGC

 101 TCAGCGCGCG TCAGCGGGTG TTGGCGGGTC TCGGGGCTGG CTTAACTATG
 AGTCCCGCGC AGTCGCCAC AACCGCCAC AGCCCGGACC GAATTGATAC

NdeI

151 CGGCATCAGA GCAGATTGTA CTGAGAGTCC ACCATATGCG GTGTGAAATA
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 201 CCGACAGAT CGCTAAGGAG AAAATACCGC ATCAGATTGG CTATTGCCCA
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 401 ACATAACTTA CCGTAAATGG CCGCGCTGGC TGACCGCCCA ACGACCCCG
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NdeI

551 GCAGTACATC AAGTGTATCA TATGCCAAGT ACGCCCCCTA TTGACGTCAA
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 601 TGACGGTAAA TGGCCCGCCT GGCATTATGC CCAGTACATG ACCTTATGGG
 ACTGCCATTT ACCGGGCGGA CCGTAATACG GGTCAATGAC TCGAATACCC

NcoI

651 ACTTCCTAC TTGGCAGTAC ATCTACGTAT TAGTCATCGC TATTACCATG
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NcoI

701 GTGATCGGGT TTTGGCAGTA CATCAATGGG CGTGGATAGC GGTTCGACTC
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EcoRV

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HindIII

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XhoI

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DraIII

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7251 CGTATCAGGA GGCCCTTTCG TC
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INTERNATIONAL SEARCH REPORT

International application No.
PCT/US99/01382

A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) : A61K 48/00; C07H 21/04; C12N 15/63, 15/86, 5/10, 15/40

US CL : Please See Extra Sheet.

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 424/450, 93.2, 93.21; 536/23.1, 23.72; 435/5, 6, 455, 457, 458, 320.1

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

APS, Dialog, Biosis, Medline, Biotech

Search terms: Ebola virus, glycoprotein, transmembrane, targeting

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	US 5,166,320 A (WU et al.) 24 November 1992, columns 9-10 and claims 1-18.	1-20
A	SANCHEZ et al. The virion glycoproteins of Ebola viruses are encoded in two reading frames and are expressed through transcriptional editing. Proceedings of the National Academy of Sciences. April 1996, Vol. 93, pages 3602-3607, especially page 3604.	3, 12
A	FELGNER et al. Lipofection: A highly efficient, lipid-mediated DNA-transfection procedure. Proceedings of the National Academy of Sciences. November 1987, Vol. 84, pages 7413-7417, especially page 7414.	1-20

☒ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

* Special categories of cited documents:	*T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
A document defining the general state of the art which is not considered to be of particular relevance	*X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
E earlier document published on or after the international filing date	*Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
L document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	*Z* document member of the same patent family
O document referring to an oral disclosure, use, exhibition or other means	
P document published prior to the international filing date but later than the priority date claimed	

Date of the actual completion of the international search 12 APRIL 1999	Date of mailing of the international search report 11 MAY 1999
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Name and mailing address of the ISA/US
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INTERNATIONAL SEARCH REPORT

International application No.
PCT/US99/01382

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	VOLCHKOV et al. GP mRNA of Ebola virus is edited by the Ebola virus polymerase and by T7 and vaccinia virus polymerases. Virology. 1995, Vol. 214, pages 421-430, especially page 424.	3, 12

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A. CLASSIFICATION OF SUBJECT MATTER:

US CL :

424/450, 93.2, 93.21; 536/23.1, 23.72; 435/5, 6, 455, 457, 458, 320.1